



ISPP INTERNATIONAL SOCIETY
FOR PLANT PATHOLOGY

PROMOTING WORLD-WIDE PLANT HEALTH AND FOOD SECURITY

INTERNATIONAL SOCIETY FOR PLANT PATHOLOGY

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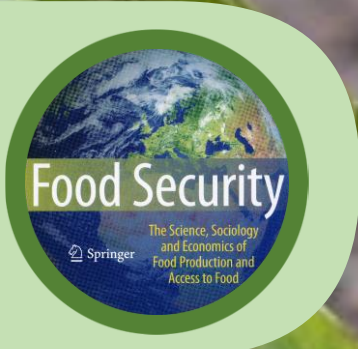
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INTERNATIONAL SOCIETY FOR PLANT PATHOLOGY (ISPP)

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THE JAKOB ERIKSSON PRIZE FOR PLANT PATHOLOGY - CALL FOR NOMINATIONS

JAKOB ERIKSSON PRIZE COMMISSION

The premier award for achievement in plant pathology, the Jakob Eriksson Prize, was established in 1923 to honor the memory of Jakob Eriksson, a prominent Swedish mycologist and plant pathologist who died in 1931. He was also a dedicated internationalist who espoused the cause of international cooperation in plant pathology. The Prize will be awarded at the International Congress of Plant Pathology held on the Gold Coast, Queensland, Australia from 19-25 August 2028 (<https://www.icpp2028.org/>). The Royal Swedish Academy of Sciences administers the Jakob Eriksson Prize Fund which provides for a gold medal award at Congresses of the International Society for Plant Pathology.

Nominations are solicited for a candidate of distinction in recognition of research in mycology, in plant pathology, or in virus diseases, or of a particular publication dealing with such subjects, with the understanding that the work being recognised is of a distinct international value and merit.

The following rules apply to those making nominations:

- i. Nominators must provide a short statement (2 pages or 500 words) justifying the selection of the nominee plus a short CV maximum three pages, and a publication list of the most relevant papers/publications or reports – maximum 20 references. Do not send a detailed Curriculum Vitae. More detail than these requirements will be sought by the Commission if required.
- ii. Names of all nominees must be strictly confidential,
- iii. Individuals cannot nominate themselves and nominators should declare any professional affiliation with the nominee.
- iv. No correspondence concerning unsuccessful nominations will be entered into.

All nominations are to be sent to the Chair of the Prize Commission, in an email headed “Jakob Eriksson Prize Nomination 2028”. Send the email to ErikssonPrize@ISPPweb.org with a c.c. to the ISPP Business Manager (andrea.masino@unito.it). The call for nominations will now close on 15 March 2027.

Prize Selection

- i. The Jakob Eriksson Prize Commission, in consultation with the Executive of ISPP, will independently undertake the selection processes to enable a recommendation of the Jakob Eriksson Prize recipient at least one year before each International Congress of Plant Pathology.
- ii. The Chair of the Commission will advise the ISPP President of the Commission’s recommendation, and after appropriate deliberation, the President of the ISPP will invite the successful nominee to accept the award.

The Prize Ceremony

- i. The participation of the Jakob Eriksson Prize recipient in the International Congress of Plant Pathology will be facilitated by the ISPP and the Congress Organisers. Normally this will include complementary Congress registration and attendance at Congress social functions, return economy travel to the Congress and some support for accommodation and reasonable expenses for the duration of the Congress.
- ii. The Prize Ceremony will be planned by the ISPP in consultation with the Prize recipient, the Commission Chair and the Congress Organisers.
- iii. As part of the Prize Ceremony, the Prize recipient will also be invited to briefly present their work at the Congress as The Jakob Eriksson Oration with scope and coverage in a style suitable for a more general audience.

The Royal Swedish Academy of Sciences will provide the Jakob Eriksson gold medal.

Information about the selection process is available here (https://www.isppweb.org/about_eriksson_2.asp).

Jacob Eriksson Prize - 1993-2023

Past recipients of the Prize are:

- 1993. 7th Recipient - Prof Dr Ir Ariena H.C. van Bruggen, Professor Biological Farming Systems at Wageningen University, at the 6th International Congress of Plant Pathology.
- 1998. 8th Recipient - Dr Richard Frederiksen, Professor of Plant Pathology at Texas A&M University, at the 7th International Congress of Plant Pathology in Edinburgh.
- 2003. 9th Recipient - Dr. Jaccov Katan of the Hebrew University, Jerusalem, at the 8th International Congress of Plant Pathology in Christchurch, New Zealand.
- 2008. 10th Recipient - Dr. Laurence V. Madden of the Ohio State University, at the 9th International Congress of Plant Pathology in Torino, Italy.
- 2013. 11th Recipient - Professor Jeffrey B. Jones of the University of Florida at the 10th International Congress of Plant Pathology in Beijing, China.
- 2018. 12th Recipient – Emeritus Professor Pierre JGM de Wit of the Laboratory of Phytopathology, Wageningen University, the Netherlands, at the 11th International Congress of Plant Pathology in Boston, USA.
- 2023. 13th Recipient – Professor Silvia Restrepo of the Universidad de los Andes, Bogota, Colombia, at the 12th International Congress of Plant Pathology in Lyon, France.



Help shape the scientific experience at ICPP 2028

Two opportunities are now open for the global plant pathology community to bring important topics, people and perspectives into the Congress experience.

BOTH PROPOSAL CALLS CLOSE

Friday 4 December 2026 | 11:59 pm AEST

Where could your scientific community make an impact?

Convene a specialist community around an extended meeting, or lead a focused session within the ICPP 2028 scientific program.

OPPORTUNITY 01

Propose a Satellite Meeting

Create a half-day, full-day or multi-day meeting for deeper scientific discussion and collaboration within a specialist community, held immediately before or after ICPP 2028.

Best for: specialist conferences, workshops and focused community meetings.



SCAN TO EXPLORE THE REQUIREMENTS

EXPLORE SATELLITE MEETINGS

OPPORTUNITY 02

Propose a Symposium Session

Shape a 90- or 120-minute session within the ICPP 2028 scientific program, combining invited expertise with research selected from submitted abstracts.

Best for: timely scientific themes suited to focused talks and moderated discussion.



SCAN TO EXPLORE THE REQUIREMENTS

EXPLORE SYMPOSIUM SESSIONS

19–25 August 2028

Gold Coast Convention and Exhibition Centre
Queensland, Australia

icpp2028.org

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Join the ICPP 2028 mailing list for conference news, speaker announcements and key deadlines.

JOIN THE MAILING LIST



SCAN TO SIGN UP

SEED PATHOLOGY WORKSHOP WITHIN BRASILIAN SEED CONGRESS - XVII BRAZILIAN SYMPOSIUM ON SEED PATHOLOGY

GIANFRANCO ROMANAZZI

Brasil is a very important country for production of seeds, and the research on seedborne pathogens is very active. The member of ISPP Seed Pathology Committee José da Cruz Machado, together with the President of Brazilian Seed Pathology Committee Norimar Denardin and other colleagues organised in Fox do Iguacu, within the 23th Brazilian Seed Congress, a Symposium on seed pathology on 27 August 2026. The theme of the Symposium was "Phytosanitary Quality of Seeds: Regulation, Traceability and Applications as a Strategic Pillar in Agricultural Production". After introducing welcome, the organisers invited Gianfranco Romanazzi, chair of ISPP Seed Pathology Committee, to provide an overview of the importance of seedborne pathogens and their management strategies, and Rose Souza Richards, ISPP Seed Pathology Committee member to discuss the effects of seedborne pathogens on international seed trade (see <https://cbsementes.com/patologia>).



Session participants (left) and social dinner (right)

Braslian Seed Pathology Committee is very active, and at the beginning of August 2025 planned a Seed Pathology session during 54th Brazilian Phytopathological Society Congress in Lavras, organised from José da Cruz Machado, with a series of talks followed on the second day by a practical application of tools to monitor seedborne pathogens in the Seed Pathology Lab of Federal University of Lavras (UFLA) (see <https://fitopatologia.com.br/programacao2>).



Seedborne pathogen session program (left) and participants to seedborne pathogen detection training (right).

MIT RESEARCHERS ENGINEER BACTERIA THAT COULD PROGRAM PLANTS FOR SELF-DEFENCE

ANNE TRAFTON, MIT NEWS, 17 AUGUST 2026

Massachusetts Institute of Technology (MIT) researchers have engineered bacteria that can function as transistors, allowing the team to create living “circuit boards” that can be printed onto a growth medium in a Petri dish. In electrical circuits, transistors function as switches that can turn current on or off. In the biological circuits that the researchers have created, bacterial switches control the flow of small molecules, which send signals to downstream circuit components.

The research team designed two different transistors, along with three bacterial strains that relay information between the transistors, giving them the building blocks they need to design nearly any type of circuit. In a new study, they used these cells to create circuits that can add two or three inputs, or send one input to a specific location in the circuit.

“We’ve built some initial computer architecture components that are commonly used, but any operation can be built with these five strains,” says Hamid Doosthosseini PhD ’25, an MIT postdoc and the lead author of the new study.

Using this approach, the researchers hope to develop circuits that one day could coat plant leaves or roots, where they could compute to sense and respond to environmental conditions such as drought or attack by pests.

Christopher Voigt, head of MIT’s Department of Biological Engineering, is the senior author of the paper, which was recently published in [*Nature Chemical Biology*](#). Former MIT postdoc Haorong Chen is also an author of the paper.

CELLS AS TRANSISTORS

When designing synthetic biology circuits, researchers typically engineer cells to express proteins and transcription factors that interact to perform a task such as sensing a target molecule, which then triggers production of a specific output.

These simple circuits can perform various logic functions, but they must use unique transcription factors to avoid crosstalk within the circuit. There is a limited number of transcription factors that can be used for these circuits, which limits the overall complexity that can be achieved in a single cell. Additionally, putting too many circuits in one cell can overburden the cell’s protein production machinery.

In the new paper, the researchers took a different approach: Instead of building an entire circuit into one cell, they designed cells that could act as transistors. These transistors can then be combined in different ways to create a variety of circuits.

To create the transistors, the researchers chose a bacterium called *Pantoea agglomerans*, which commonly grows on surfaces, including plants. Using these cells, they made two types of transistors that can be switched on or off by a molecule called OC-6. One of the transistors is switched on by this input, and the other is switched off. Each transistor also detects the presence of a target molecule, in this case, OC-12. Depending on whether that molecule is present, and whether the switch is active, the transistors produce an output molecule known as OHC-14.

The researchers also used three strains of *Pantoea agglomerans* to create relays, which translate the OHC-14 signal into an output that can be fed into another transistor. Using these relay strains, the researchers can “wire” the transistors together, just like an electronic circuit board.

For example, they could create a bidirectional switch with two transistors that sense OC-12, and then send that information to different relay strains based on a switch input, ultimately feeding into other transistors that further process the signal.

The researchers created their circuits by printing colonies of bacteria onto plates containing agar, a growth medium. Each colony is printed about 5 millimeters from the nearest one. This allows the signals to travel only to the nearest colony, which then relays them to the next one, so information flows only in one direction.

COMPLEX CALCULATIONS

In this paper, the researchers demonstrated a transistor that can perform several types of logic operations depending on its location in the circuit layout, including “multi-input,” “or,” and “imply” gates. They also combined the transistors to create more complex circuits that can add up two signals, process more signals simultaneously, or function as a demultiplexer — a circuit that takes one incoming signal and sends it to one of several possible destinations, depending on a control signal.

The largest of these circuits, which adds two inputs together, contains 24 bacterial colonies wired together.

“This work shows that we can get toward more complicated functions by linking up simpler functions in individual cells,” Voigt says. “Computationally, there’s nothing that your iPhone can do that these circuits couldn’t do.”

Circuits made from these cells take about eight hours to perform each calculation, much longer than a computer circuit. But, for biological applications, that is a reasonable amount of time, the researchers say.

“We’re not trying to replace computers, but rather put computational control into biology. If you have bacteria on the root of a plant, or the plant itself is doing the computing, running a simple calculation overnight is fast enough relative to a growth season,” Voigt says.

If developed for use in agriculture, this type of circuit could be applied to the roots of plants to detect different types of stress. Once a particular input is detected, it would trigger a response such as synthesizing a fungicide.



RESEARCHERS CREATE NEW WEAPON AGAINST TOMATO SPOTTED WILT

JAMIE MARTIN, WISCONSIN AG CONNECTION, 2 OCTOBER 2026

A new research breakthrough from Texas A&M AgriLife Research could help plant breeders develop peppers, tomatoes and other crops with stronger resistance to tomato spotted wilt virus (TSWV). The technology was developed by a team led by Jeanmarie Verchot, Ph.D., professor in the Texas A&M Department of Plant Pathology and Microbiology. The researchers developed the first infectious clone of TSWV produced by U.S. scientists, providing a standardised research tool for studying the disease and the plant genetics associated with resistance.

Their work was detailed in the paper, "[A Novel Approach to Engineering Tomato Spotted Wilt Virus Infectious Clones by Disarming Key Nodes in Anti-Viral Defenses](#)," recently published in *Molecular Plant-Microbe Interactions* by the American Phytopathological Society.

An infectious clone allows researchers to reproduce the virus in a controlled research environment. According to the Texas A&M team, the technology makes it possible to generate an unlimited supply of virus and inoculate plants used in breeding trials more efficiently. This could reduce the time breeders need to identify pepper and tomato genetics that can resist TSWV.

"Through our research, we can now produce an unlimited amount of virus, more efficiently inoculate trial plants and help pepper breeders move much faster in identifying superior, virus-resistant genetics," Verchot said.

TSWV is a serious agricultural disease that affects more than 1,000 produce and ornamental plant varieties around the world. It is transmitted by thrips, insects that can spread the virus between plants and move into agricultural areas through regional wind patterns.

Disease infection can reduce crop yields and cause visible damage that makes produce unsuitable for sale. For growers, those losses can reduce income while increasing production costs. Consumers may ultimately face higher prices when disease reduces the available supply of produce.

The USDA Risk Avoidance and Mitigation Program estimates that TSWV results in about \$100 million in crop damage each year in Florida, Georgia, North Carolina and South Carolina.

Growers have traditionally relied on crop varieties with genetic resistance to TSWV. However, new virus strains are emerging that can overcome existing resistance. Scientists have been examining the genomes of these strains to understand why some cause more severe disease.

The Texas A&M researchers developed a full-length complementary DNA copy of the genome from a TSWV strain that infects peppers. The clone produces infectious virus similar to the original isolate and provides researchers with a consistent starting point for experiments.

"This clone allows all U.S. researchers the same starting point, providing a more efficient tool for identifying the virus in crops and for creating disease management practices," said Haden Ball, Verchot's graduate student.

DISCOVERING A NEW DIAGNOSTIC TOOL FOR DETECTING HARMFUL BACTERIA

KATIE BOHN, PENN STATE NEWS, 15 SEPTEMBER 2026

Scientists, clinicians, seed companies and others may soon have a new tool in their belt for detecting bacterial pathogens, thanks to a new study by researchers in Penn State's College of Agricultural Sciences. The research in the *Journal of Microbiological Methods* details a new method for detecting *Pseudomonas syringae*, common bacteria that infect a wide variety of crops, vegetables and woody ornamentals.

The cost is similar to the traditional method, called enzyme-linked immunosorbent assay or ELISA. However, the researchers' new method — called enzyme-linked chaperone assay or ELCA — has the potential to detect only living bacteria, making it more sensitive and accurate.

Rachel Herschlag, lead author on the paper who earned her doctorate in plant pathology at Penn State, said the study is a proof of concept, with opportunity in the future to adapt the method for detecting other pathogens. "Now that we've documented this method, researchers can adapt it for other bacteria that have a similar system," she said. "This could include some animal and human pathogens in addition to those affecting plants, expanding its potential utility in clinical, agricultural, environmental and biotechnology applications."

Carolee Bull, professor of bacterial systematics and plant pathology and co-author of the paper, said that many current diagnostic tests like ELISA work by using animal-derived antibodies produced to fight an infection. However, the new method uses a different mechanism. "I asked Dr. Herschlag to think creatively about bacterial pathogen detection to develop novel strategies for detecting *Pseudomonas* strains, and potentially, other types of bacteria," Bull said. "She went on to develop this technique by using the same mechanism that many pathogens use to invade their hosts to allow us to recognise them."

When bacteria attack plants, animals or people, they deliver specialised proteins called effectors into the host's cells, she explained. These proteins help the bacteria overcome the host's immune system and invade the organism. To do this efficiently, many bacteria rely on helper proteins called chaperones that recognise and bind — like a lock and key — to these effectors with high specificity. The chaperones protect the effectors, which are vulnerable to degradation.

The new method relies on this process, rather than the labor-intensive process of harvesting antibodies in a lab. "Different from other diagnostics, this new method uses a bacterial chaperone to recognize and bind to its matching effector protein," Herschlag said. "This then triggers a colorless solution to turn yellow, providing an easy visual signal that the target is present. And because this interaction is highly specific, the method can reliably detect its intended target."

Other diagnostic approaches, the researchers explained, work by detecting specific parts of the cell such as DNA. However, since these parts exist even after bacteria have died, this can produce false positives. The researchers said what's most helpful about this new technique is that effectors are only produced by living bacteria.

“Because effectors are thought to degrade quickly, effectors from dead bacteria likely do not survive very long,” Herschlag said. “Thus, ELCA can detect living bacteria, or in other words, those that are actually capable of causing infections. Indeed, we showed that living bacteria had higher levels of detection than dead bacteria.”

In the future, the researchers said next steps will be to optimise the test by reducing background signal, along with adapting the concept for other pathogens that have comparable chaperone-effector systems, including some that are relevant to agriculture, veterinary science and human health.

HOW A PATHOGEN TURNS FLOWERS INTO LEAVES

THE EUROPEAN SYNCHROTRON NEWS, 16 SEPTEMBER 2026

Scientists have found how a protein produced by bacteria disrupts the mechanism that controls flower development in plants, turning reproductive organs into leaf-like structures. These findings shed light on a plant disease that is expected to spread as climate change expands the range of its insect vectors. They used ESRF's beamline BM29. The results are the Editor's Pick in the *Journal of Biological Chemistry*.

Phytoplasmas are bacteria that are transmitted to plants by insects. The symptoms of the pathogen can range from triggering dwarfism in the plant to essentially reprogramming plant development: instead of flowers, all the floral organs- sepals, petals, stamen and carpels- are converted to leaves, with diverse crops such as sunflowers, sesame and grapevine affected.



This effect can have economic consequences, as food might not be as easily available in damaged crops. For example, outbreaks of phytoplasma coconut lethal yellowing disease in Africa since the 1930s has led to devastating crop losses up to 40% in Tanzania and 20% in Ghana. In addition, climate change has increased the presence of insects in more northern latitudes, which increases the presence of phytoplasmas. “Understanding how they hijack the plant’s own developmental machinery could help us find strategies to hopefully mitigate their impact on agriculture”, explains Chloe Zubieta, CEA researcher at the Laboratoire de Physiologie Cellulaire et Végétale, ESRF visiting scientist and leader of the study.

Zubieta joined forces with ESRF scientists Mark Tully and Max Nanao to study a phytoplasma protein called PHYLOY (phylogen protein from *Candidatus Phytoplasma asteris*, onion yellows strain), which interferes with the plant’s normal development. It does this by targeting plant proteins called MADS-box transcription factors (MTFs).

MTFs play an important role in deciding how a flower develops. They work together in groups of four, binding to DNA and switching on the genes that tell the plant which parts of the flower to produce.

The researchers used Small X-ray scattering on ESRF’s beamline BM29 to study how PHYLOY interacts with three important MTFs. Their experiments showed that PHYLOY can interact with a wider range of MTF combinations than previously thought. “We can observe flexibility in the SAXS data and we think this corresponds to the interaction plasticity of phylogen and host plant transcription factors,” notes Mark Tully, beamline operation manager at BM29.

The researchers then changed individual parts of these proteins using a technique called site directed mutagenesis and found that they could either prevent or enable the interactions. This showed that PHYLOY recognises specific features shared by different MTFs, allowing the pathogen to interfere with the plant’s developmental machinery by structural mimicry.

By preventing MTF proteins from working together properly, the phytoplasma protein changes the identity of floral organs, causing them to develop into leaf-like structures.

“We are working on obtaining the high-resolution crystal structure of the MTF-PHYLOY complex and hope to use these data, in combination with high throughput virtual screening, to design inhibitors of this interaction,” comments Max Nanao, Deputy Group Head of MXIS.

DETECTING CHESTNUT INK DISEASES THROUGH THE DOG'S SENSE OF SMELL: FROM LABORATORY TO FOREST

A paper by Francesca Carloni *et al.* titled “Detecting Chestnut Ink Diseases through the dog's sense of smell: From laboratory to forest” was published on 7 September 2026 by *Plant Pathology* (Vol. 75, Issue 5, e70269). The abstract is as follows:-

Chestnut ink disease, caused by *Phytophthora* spp., represents one of the main threats to the productivity and sustainability of sweet chestnut (*Castanea sativa*) in southern Europe. Early diagnosis is an essential step to develop appropriate management strategies. Therefore, this study investigates the potential of dogs' olfactory abilities to detect pathogen-specific volatile organic compounds (VOCs) emitted by infected roots, offering a promising, non-invasive approach for rapid and reliable disease detection in forest ecosystems. Hulk, a 7-year-old Labrador retriever, was trained through a multiphase protocol using GETXent polymer tubes soaked with *Phytophthora* VOCs under controlled conditions. His performance was then evaluated in greenhouses with artificially inoculated seedlings and in chestnut orchards under natural conditions. Statistical analyses showed that Hulk demonstrated positive *Phytophthora* detection skills throughout the training phases, both with polymer tubes and in the greenhouse and his performance remained consistently superior to random expectations. In chestnut orchards, Hulk revealed high sensitivity, showing signs of alert for 86% of trees displaying specific ink disease symptoms and for 75% of trees displaying nonspecific symptoms. Furthermore, the dog was shown to be efficient in intercepting early root infection on apparently asymptomatic trees, validated by laboratory analyses. In addition, VOCs related to *Phytophthora* infections were chemically identified as o-cymene, camphor and borneol. To our knowledge, this is the first application of canine detection using GETXent polymer tubes to train a dog for *Phytophthora cambivora* surveillance in chestnut orchards, which could have application in nursery stock to limit anthropogenic spread on a wide scale.

[Read paper.](#)

CURRENT VACANCIES

Assistant Professor of Plant Pathology – Plant Health and Microbiomes, University of California, Davis, USA

The Department of Plant Pathology in the College of Agricultural and Environmental Sciences at the University of California, Davis is recruiting a tenure-track Assistant Professor of Plant Pathology, with an emphasis on plant health and microbiomes. The academic-year (9-month) appointment includes a role in the California Agricultural Experiment Station. The salary range is US\$106,800–130,700 per year.

The successful candidate is expected to establish a competitive, externally funded research program in plant microbiome science addressing critical and emerging issues in plant pathology. Research should advance microbiome-informed approaches to the diagnosis, prediction and/or management of plant diseases and may include disease-suppressive microbiomes, pathobiomes, host–microbiome–pathogen interactions, disease complexes, microbiome engineering, synthetic communities, and climate-driven changes in plant–microbiome interactions.

The appointee will contribute to undergraduate and graduate teaching, including courses in microbiomes, Global Disease Biology, One Health, plant pathogen ecology and epidemiology, and microbial ecology. Graduate and undergraduate mentoring, outreach, and departmental and university service are also expected.

More information and applications:

https://www.isppweb.org/Ads/PlantPathology_AssistantProfessor_CaliforniaDavis.pdf.

ACKNOWLEDGEMENTS

Thanks to Grahame Jackson, Greg Johnson, Jan Leach, and Gianfranco Romanazzi for contributions.

COMING EVENTS

7th International Symposium on Fusarium Head Blight

5 October – 8 October, 2026

Department of Agricultural, Food and Environmental Sciences, University of Perugia

Perugia, Italy

Website: www.7isfhb.org

International Phytobiomes Conference 2026

3 November – 5 November, 2026

Niagara-on-the-Lake, Ontario, Canada

Website: <https://phytobiomesconference.org/>

International Plant and Animal Genome Meetings (PAG34)

8 January – 12 January, 2027

San Diego, California, USA

Website: <https://intlpag.org/PAG/>

XXI International Plant Protection Congress (IPPC) 2027 in conjunction with 26th Australasian Plant Pathology Conference (APPC)

1 November – 5 November, 2027

Te Pae Christchurch Convention Centre, Christchurch, New Zealand

Website: www.ippc2027.com

13th International Congress of Plant Pathology 2028

19 August – 25 August, 2028

Gold Coast, Queensland, Australia

Website: www.icpp2028.org



ICPP 2028 13th International Congress of Plant Pathology
19-25 August, Gold Coast Convention & Exhibition Centre, Queensland, Australia

INTERNATIONAL SOCIETY FOR PLANT PATHOLOGY (ISPP)

WWW.ISPPWEB.ORG

The ISPP List is an e-mail list server which broadcasts messages and announcements to its subscribers. Its goal is to facilitate communication among members of the International Society for Plant Pathology and its Associated Societies. Advertised vacancies in plant pathology and ISPP Newsletter alerts are also sent to members of the ISPP List.

In accordance with the guidelines and recommendations established by the new EU General Data Protection Regulation 679/2016 (GDPR), the International Society for Plant Pathology has created a [Privacy Information Notice](#) containing all the information you need to know about how we collect, use and protect your personal data.

This policy explains when and why we collect personal information about our users, how we use it, the conditions under which we may disclose it to third parties, how we keep it safe and secure and your rights and choices in relation to your personal information.

Should you need further information please contact business.manager@issppweb.org

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