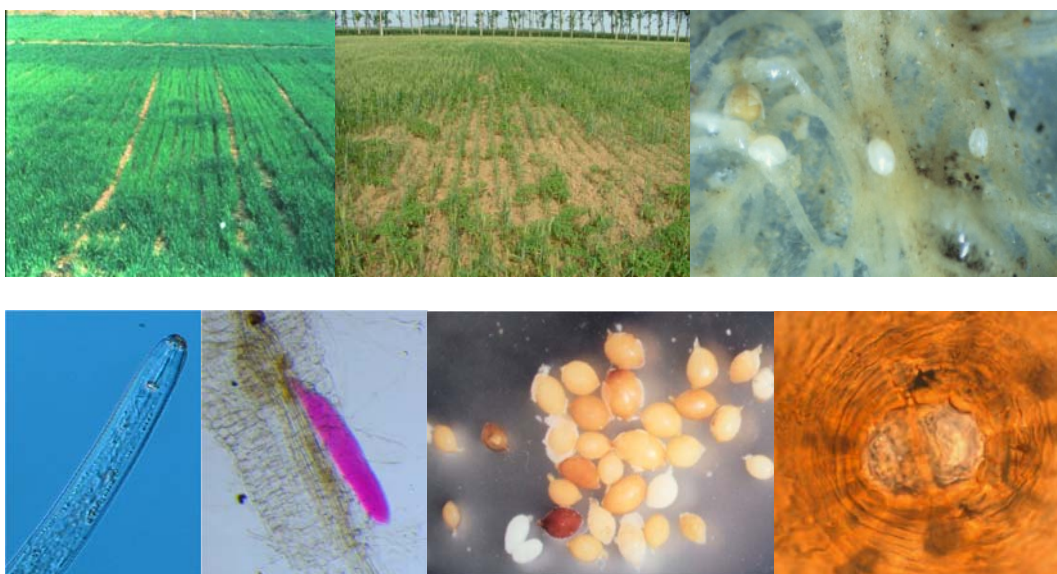


The 4th International Cereal Nematodes Initiative Workshop

第四届国际禾谷类线虫病害学术研讨会

August 22-24, 2013

Beijing·China



Sponsored by

Institute of Plant Protection, Chinese Academy of Agricultural
Sciences

International Maize and Wheat Improvement Center (CIMMYT)
China Agricultural University

State Key Laboratory of Biology of Plant Diseases and Insect Pests
Chinese Society for Plant Pathology
Chinese Society of Plant Nematologists



CONTENTS

General Information	-----1
Scientific Program	-----5
Abstracts of Scientific Program	-----11
General Abstracts	-----48
Participant Lists	-----55
Volunteers	-----59
Map of Friendship Hotel and Beijing	-----60

General Information

Letter of Invitation

Dear Friends & Colleagues,

It is our pleasure to invite you and your collaborators to attend the 4th International Cereal Nematodes Initiative Workshop (ICNI) in Beijing, China. This event, which will take place on 22nd-24th August 2013, and provide a stimulating platform for exchanging research developments and tracking technical progress on cereal nematodes research.

The 4th ICNI is a Pre-workshop included in the program of the 10th International Congress of Plant Pathology (ICPP), to be held in Beijing from 25th to 31st August 2013, thus allowing participants to attend both 4th ICNI and ICPP minimizing time and costs.

The 4th ICNI has been jointly sponsored by the Institute of Plant Protection, Chinese Academy of Agricultural Sciences (IPP-CAAS), CIMMYT International Maize and Wheat Improvement Center, China Agricultural University and is supported by the State Key Laboratory of Biology of Plant Diseases and Insect Pests (SKLBPI), and the Chinese Society for Plant Pathology (CSPP).

We sincerely invite you to come together with us, make this workshop enjoyable and profitable.

We wish you have a good stay in Beijing.

On behalf of the 4th ICNI organizing committee

Prof. Deliang Peng and Dr. Francesco Di Serio

Organizers

Dr. Deliang Peng, Institute of Plant Protection, Chinese Academy of Science, China, Email: dlpeng@caas.net.cn

Dr. Amer A. Dababat, CIMMYT, Ankara, Turkey, Email: a.dababat@cgiar.org

Scientific Committee

Dr. Deliang Peng – IPPCAAS, China, Email: dlpeng@caas.net.cn

Dr. Amer A Dababat, CIMMYT, Ankara, TURKEY, Email: a.dababat@cgiar.org

Dr. Julie Nicol – CIMMYT International, Email: j.nicol@cgiar.org (Chair ICCNI)

Prof. Maurice Moens – ILPO, Belgium, Email: maurice.moens@ilvo.vlaanderen.be

Prof. John Jones – SCRI, UK, Email: john.jones@scri.ac.uk

Dr. Richard Smiley – OSU, USA, Email: richard.smiley@oregonstate.edu

Dr Zahra Tahna Maafi – Iran, Email: tanhamaafi@yahoo.com

Prof Halil Elekcioğlu – Cukurova University, Turkey, Email: halile@cu.edu.tr

Local Organizing Committee

Dr. Wenkun Huang (China)

Dr. Lingan Kong (China)

Prof. Heng Jian (China)

Prof. Jinling Liao (China)

Prof. Yuxi Duan (China)

Prof. Julian Chen (China)

Ms. Wenting He (China)

Prof. Shulong Chen (China)

Associate Prof. Yanhua Wen (China)

Prof. Jingwu Zheng (China)

Prof. Hongmei Li (China)

Prof. Xianqi Hu (China)

Workshop Topics

1. Global status of the distribution of cereals nematode (CN)
2. The economic importance and population dynamics of CN on wheat
3. Control strategies of CN in wheat using host resistance
4. Control strategies of CN other than host resistance
5. Use of molecular tools for research with CN (such as pathogen diagnostics, phylogeny studies and host resistance).
6. CN genome and parasitism genes
7. Biological control and IPM

Registration and Abstract Submission

Registration and abstract submission of this workshop are bundled with the 10th ICPP registration system.

Early bird registration before Jan. 15, 2013: RMB 1,600; Onsite registration: RMB 2,100

Official published proceedings will be made from this workshop. If you would like to contribute to the workshop please kindly submit your abstract of no more than 250 words via 10th ICPP web and also email to Dr. Deliang Peng (dlpeng@ippcaas.cn) and Dr. Amer Dababat (a.dababat@cgiar.org) and the scientific committee for the workshop will review applications and inform contributors of acceptance as poster/paper by 1 June 2013. Contributors with accepted papers will be asked to prepare a 2-4 page paper by June 16, 2013. Please follow the format provided below.

Accommodation

Hotel costs are RMB 490/single room at Friendship Hotel Grand. The hotel is located 40 minutes away from airport by car and can be reached with Shuttle Bus for about 16 RMB. Please contact workshop organizer Deliang Peng at dlpeng@caas.net.cn; phone +86-10-62815611; fax +86-10-62896114 for hotel reservations.

Weather

August in Beijing is sunny, warm and humid. The average daily temperature is 25-33°C (77-95°F) .

Insurance

The registration fees do not include insurance for the participants regarding accidents, sickness or loss of personal property. You are advised to make necessary arrangements for a short-term health and accident insurance before leaving your home country.

Currency Exchange

In China, only RMB is used. Exchange centers can be found at airports, hotels. The exchange rate is set by the Bank of China, which is now about US \$1.00=RMB 6.15. When exchanging money, you should keep your receipt in case you want to change any RMB back to foreign currency when leaving China.

Credit cards (Visa, Master, American Express, Diners Club and JCB) are accepted in many department stores and hotels. It is possible to draw cash from ATM machine by above credit cards.

The Bank of China and most of the hotels can cash traveler cheques issued by most foreign banks or financial institutions.

Arrival in Beijing

From Beijing Capital International Airport to Friendship Hotel Grand. The hotel is located 40 minutes away from airport by car and can be reached with Shuttle Bus for about 16 RMB. It takes about 30 mins by taxi, and the charge is around RMB 100 for one way.

Most important: Please print the following picture and show it to TAXI driver, who may deliver you to the Friendship Hotel Grand.



Check-in

There will be reception staff to help you check-in at 8:00 am-22:00 pm on August 22, at the Building 2, Jingbing Lou, Friendship Hotel Grand.

For those who have not paid registration fee online, you will need to pay it on-site.

After checking-in, you will get an information pack, meal vouchers, and workshop documents.

On-site registration cash (RMB) only, 2100 RMB for each delegate and 600 RMB for each family member.

Please you in advance change to RMB at the airport or Friendship Hotel Grand.

Preliminary Schedule

8:00 - 20:00, August 22, 2013 Yinbinlou, Friendship Hotel Grand, Registration;
8:30 - 19:00, August 23, 2013 Jiabinlou, Friendship Hotel Grand, Scientific program;
19:30 - 22:00, August 23 2013 Welcome reception and banquet ,Yuegongfu, Xueyuan South Road;
8:30 - 12:30, August 24, 2013 Jiabinlou, Friendship Hotel Grand, Scientific program;
14:00- August 24, 2013 Check out, Workshop end

Scientific Program

The conference will be held in August 23rd-24th, 2013, in Beijing. The conference covers a wide range of topics on cereal nematodes.

All meetings will take place in the first Meeting Room at Friendship Hotel Grand, Yibing Lou, on August 23-24, 2013.

For the speakers, please copy your PPT files to the computer between 14:00 pm and 22:00 pm on August 22 on Registration Site at Friendship Hotel Grand, Jingbin Lou or 7:30 am-8:20 am and 13:00-13:30 pm on August 23-24 at meeting room.

Agenda for the 4th International Cereal Nematodes Initiative Workshop
August 22-25, 2013, Beijing, China

Organizers: Prof. Deliang PENG & Dr. Amer DABABAT	
August 22, 2013	Registration: Lobby of Building No 2(Jingbinlou)in Friendship Hotel, Beijing(北京友谊宾馆 3 号楼, 敬宾楼大厅) Drs. Wenkun HUANG & Ligan KONG
August 23, 2013	Building 8, Jiabinlou, First Meeting Room(8 号楼, 嘉宾楼第一会议室)
8:30- 9:10	Opening Ceremony, Photos
SESSION I : 9:10-10:30	Global status of the distribution of cereal nematodes: Chairman: Prof. Halil ELEKCIOĞLU Prof. Heng JIAN
9:10-9:30	1. Amer DABABAT: Cereal nematodes management strategies in wheat
9:30-9:50	2. Richard W. SMILEY: Resistance and tolerance to <i>Heterodera avenae</i> in North American spring wheat
9:50-10:10	3. Deliang PENG: Current research status and perspective of cereal cyst nematode, (<i>Heterodera aveane</i> , <i>H. filipjevi</i>) on wheat in China
10:10-10:30	4. Zahra Tanha MAAFI: Cereal cyst nematodes and its research perspectives in Iran
10:30-10:45	Tea Break
SESSION II 10:45-12:00	The economic importance and biology of cereal nematodes Chairman: Dr. Amer DABABAT, Prof. Jinling LIAO
10:45-11:00	5. Shulong CHEN: The biology of cereal cyst nematode in Hebei Province, China: implications on its control
11:00-11:15	6. Alireza AHMADI: Crop loss assessment of <i>Heterodera filipjevi</i> on some cultivars of wheat, barley and triticale under field condition of Southwest of Iran
11:15-11:30	7. Ricardo HOLGADO: Importance of identification cereal cyst nematodes species and pathotypes in breeding for resistance and for management strategies
11:30-11:45	8. Jie ZHANG: The effect of the cereal cyst nematode, <i>Heterodera avenae</i> , on wheat yield in Shangdong province, China
11:45-12:00	9. Mustafa İMREN: Studies on cereal cyst nematodes, <i>Heterodera avenae</i> Wollenweber, in South East Anatolia and Eastern Mediterranean Regions in Turkey
12:00-13:30	Lunch
SESSION II 13:30 -14:30	The economic importance and biology of cereal nematodes Chairman: Dr. Zahra Tanha MAAFI, Dr. Shulong CHEN
13:30-13:45	10. M.M.M. ABD-ELGAWAD: Spatial distribution of the cereal cyst nematodes in Egyptian bread wheat (<i>Triticum aestivum</i> cv. Giza 68) field

13:45-14:00	11. Hongxia YUAN: Field dynamics and hatching characteristics of two species of wheat cereal cyst nematode in Henan province, China
14:00-14:15	12. Honghai ZHAO: The effect of the rotation with peanut on CCN population and wheat yield
14:15-14:30	13. Fouad. MOKRINI: Morphometrical and molecular characterization of root lesion nematodes (<i>Pratylenchus</i> spp.) on wheat in Morocco
Session III 14:30-15:45	Control strategies of CCN in wheat using host resistance Chairmen: Prof. Richard W. SMILEY, Prof. Hongmei LI
14:30-14:45	14. Halil ELEKCIOĞLU: Screening of international spring wheat germplasms against the cereal cyst nematode (<i>Heterodera avenae</i>) and the root lesion nematodes (<i>Pratylenchus thornei</i> and <i>P. neglectus</i>) in-vitro conditions in 2012, Turkey
14:45-15:00	15. Hongjie LI: Toward management of cereal cyst nematode with host resistance: identification of effective resistant sources in China
15:00-15:15	16. Halil TOKTAY: New sources of resistance against the root lesion nematodes, <i>Pratylenchus thornei</i> Sher et Allen in some national and international wheat germplasm
15:15-15:30	17. Andreas WESTPHAL: Identifying a novel source of resistance against <i>Heterodera filipjevi</i> in spring barley ‘Steptoe’ by analysis of a double haploid population Steptoe x Morex
15:30-15:45	18. Kh.A MOUSTAFA: Selection of certain wheat genotypes for resistance to cereal cyst nematode (<i>Heterodera avenae</i>) based on growth parameters and molecular markers
15:45-16:00	Tea Break
Session IV 16:00-17:00	Control strategies of CCN other than host resistance Chairmen: Dr. Ricardo HOLGADO, Dr. Yuxi DUAN
16:00-16:15	19. Richard W. SMILEY: Influence of abamectin and <i>Bacillus firmus</i> on <i>Heterodera avenae</i> and on spring wheat yields in the USA
16:15-16:30	20. M.M.M. Abd-Elgawad: Effect of essential oils of some medicinal plants on phytonematodes in Egypt
16:30-16:45	21. Al-Hazmi: <i>Verticillium chlamydosporium</i> , a fungal parasite of the cereal cyst nematode (<i>Heterodera avenae</i>) in the Saudi fields
16:45-17:00	22. Hongmei LI: Toward integrated management of cereal cyst nematodes in China: an example for research work in Jiangsu Province
Session V 17:00-17:45	Molecular diagnosis of cereal cyst nematodes Chairmen: Prof. Andreas WESTPHAL, Prof. Yanhua Wen
17:00-17:15	23. Gaofeng WANG: Sensitive and direct detection of <i>Heterodera filipjevi</i> in soil and infected wheat by species-specific SCAR-PCR assays
17:15-17:30	24. Fateh TOUMI: Quantitative detection of <i>H. avenae</i> and <i>H. filipjevi</i> using qPCR
17:30-17:45	25. Guiping YAN: Developing species-specific PCR assays for identification of <i>Heterodera filipjevi</i> and <i>H. avenae</i>

18:00-21:00	Banquet Yuegongfu (粤公府), Xueyuan South Road No.59, 学院南路 59 号 (农科院南门向东 100 米,魏公村路口东 500 米) Contact Dr. Wenkun HUANG
August 24, 2013	Buliding 8, Jiabinlou, First Meeting Room (8 号楼, 嘉宾楼第一会议室)
Session VI 8:30-9:30	Genome and parasitism genes Chairmen: Prof. Mike JONES, Dr. Deliang PENG
8:30-8:45	26. Mike JONES: Gene silencing in root lesion nematodes significantly reduces reproduction in host plants
8:45-9:00	27. Joong-Ki PARK: Comparative analysis of complete mitochondrial genome sequences confirms independent origins of plant-parasitic nematodes
9:00-9:15	28. Dan YANG: Comparison of transcriptome pre- and post-parasitic stages of the nematode <i>Heterodera avenae</i>
9:15-9:30	29. Changlong CHEN: Identification of an annexin-like parasitism gene from cereal cyst nematode, <i>Heterodera avenae</i>
9:30-9:45	Tea Break
Session VII 9:45-11:45	Biological control and IPM Chairmen: Prof. Joong-Ki PARK, Prof. Jingwu ZHENG
9:45-10:00	30. Amer DABABAT: Control options against the cereal cyst nematodes <i>Heterodera filipjevi</i> in Turkey
10:00-10:15	31. Haiyan WU: Occurrence and Development of the Cereal Cyst Nematode (<i>Heterodera avenae</i>) in Shandong, China
10:15-10:30	32. Yuxi DUAN: The researches of <i>Heterodera glycines</i> and resistant soybean germplasm in China
10:30-10:45	33. Jie ZHANG: Biocontrol of cereal cyst nematode by two bacteria agents
10:45-11:00	34. Kan ZHUO: First report of cystoid nematode in China with notes on <i>Cryphodera sinensis</i> n. sp.(Nematoda: Heteroderidae)
11:00-11:15	35. Hai LONG: Diagnosis of <i>Cryphodera brinkmani</i> intercepted on <i>Juniperus chinensis</i> imported into China from Thailand
11:15-11:30	36. Xiangxia BU: Temperature-manipulated development of novel cellular and mycelium stages of <i>Pasteuria penetrans</i>
11:30-11:45	37. Congli WANG: QTL analysis of gene RKN2 in <i>Gossypium barbadense</i> which clusters with gene rkn1 in <i>G. hirsutum</i> for transgressive nematode resistance
11:45-12:00	Closing remarks
12:00-14:00	Lunch
14:00	Workshop end

Abstracts of Scientific Program

SESSION I : Global status of the distribution of cereal nematodes

August 23, 9:10-10:30

Chairman: Dr. Prof. Halil ELEKCIOĞLU

1 Cereal nematodes management strategies in wheat

A.A. Dababat, G. Erginbas-Orakci, H.J. Braun, and A. Morgounov

Amer Dababat: CIMMYT Global Wheat Program, Ankara-Turkey,

Corresponding author: a.dababat@cgiar.org

Soil borne pathogens (SBPs) including the Cereal Cyst Nematode (CCN) caused by *Heterodera* species and the Root Lesion Nematodes caused by *Pratylenchus* species are attack roots of cereal crops and resulting in a high yield loss and reduce grain quality. The damage caused by these nematodes is accelerated in areas where drought exists. A few control options are being used to reduce CCN damage through keeping the population level below damage threshold such as; chemical, biological, cultural, and genetic (resistance/tolerance) practices. Resistance is environmentally friendly and biologically effective once identified. However, up to now, resistance has only been identified against one of the CCN nematodes, *Heterodera filipjevi* in Turkey and foreign wheat germplasm though this resistance is not yet present in high yielding cultivars. Resistance to the other nematodes in the CCNs complex is still being sought. Therefore, alternative approaches limiting the damage caused by CN to wheat are needed. As a result of screening wheat germplasm against the CN hundreds of moderately resistant germplasm to *H. filipjevi* in winter wheat and to both *Pratylenchus* species in spring wheat germplasm are available. In 2012, germplasm with multi disease resistance including *H. avenae*, *Pratylenchus thornei* and *P. neglectus*, and *H. filipjevi* were distributed to international collaborators. The preliminary results of using seed treatments showed that seed treatment of wheat susceptible germplasm gave up to 47% reduction in number of *Heterodera filipjevi* cyst per plant but did not reduce the number of cyst in the resistant germplasm since the cyst number was low and no room to decrease it further.

2 Resistance and tolerance to *Heterodera avenae* in North American spring wheat

R.W. Smiley, J.M. Marshall and G.P. Yan

Oregon State University, Pendleton, Oregon, 97801, U.S.A.

Corresponding author: richard.smiley@oregonstate.edu

The cereal cyst nematode, *Heterodera avenae*, reduces wheat production efficiency by >US\$3.4 million annually in the Pacific Northwest states of Idaho, Oregon and Washington. Spring wheat trials were conducted in naturally-infested fields in Idaho and Washington during 2012. Twenty cultivars were planted as a split-plot design with each cultivar planted into six replicates of 1.8 × 9-m plots that were either treated with nematicides or were untreated. All plants exhibited the typical root knotting symptom in untreated soil. Mean number of white females was greater ($P < 0.01$) in untreated (9/plant) than in treated soil (<1/plant). In untreated soil, fewer white females were produced on Ouyen and WB-Rockland (1/plant) than on other cultivars (9-29/plant). Post-harvest density of *H. avenae* eggs (from cysts) was higher (>12,000/kg of soil) following growth of susceptible cultivars than following Ouyen and WB Rockland (<5,000/kg of soil). The latter density was similar to that in plots of all cultivars produced in treated soil, indicating a natural background density of *H. avenae* remaining from cysts that had been produced on cereal crops one or two years earlier. Ouyen contains *Cre1* resistance but was intolerant, with grain yield being lower in untreated than in treated plots. WB-Rockland was both highly resistant and tolerant. Two cultivars (Buck Pronto and UI Stone) were highly susceptible but very tolerant. These were the first field trials in North America that demonstrated the benefits that can be expected from developing cultivars with resistance plus tolerance to *H. avenae*.

3 Current Research status and perspective of cereal cyst nematode, (*Heterodera avenae*, *H. filipjevi*) on wheat in China

Deliang PENG, Wenkun Huang, Huan Peng

State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China.

Corresponding author:: pengdeliang@caas.cn

The cereal cyst nematode (CCN), *Heterodera avenae* is one of the most important plant parasitic nematode and distributes throughout nearly all cereal growing areas in China. Since 2009, The occurrence and distribution of the cereal cyst nematodes (*H. avenae*) has been confirmed to occurrence in four new provinces including Ningxia, Tianjing, Xizang (Tibet) and Xijiang based on the morphological identification and molecular characterisation, so CCN has been confirmed to distribute 16 provinces in China. Yield losses were tested with inoculation and field, in Henan, Hebei, Beijing suburb, Qinghai, yield losses reach up to 18%-35% in Henan, 15-20% in Hebei, 11-18% in Beijing Suburb and 10%-28.24% in Qinghai. The population dynamics and life cycle of *H. avenae* were investigated in Beijing, Hebei, Jiangsu and Shandong from 2010 December to 2012 December. The results showed that there is one generation of *H. avenae* per year in those area above mentioned. There are two cyst nematodes (*H. avenae* and *H. filipjevi*) occurrence in the wheat production area of China, the *H. avenae* is dominate species. The molecular diagnosis methods based on SCAR-PCR were developed to identify and early detect *H. avenae* and *H. filipjevi* from infested field. The pathotypes of twenties population of *H. avenae* from Beijing, Hebei, Jiangsu and Shandong were tested and identified by using the International Test Assortment (CIMMYT provide). The results showed that Qingyundian population (*H. avenae*) of Beijing was different from the 13 pathotypes which have been described and nominated. It's slightly similar with pathotype Ha23. More than two hundred cultivars were tested and evaluated resistance to *H. avenae* and *H. filipjevi* in greenhouse and the field respectively. The results showed that no immune materials were found. Five cultivars including VP1620, BATAVIA, SUNR23, AUS4930 6.5/GS50a and Taikong 6 were high resistance to CCN Beijing population.

A cDNA library from the second stage juveniles of *H. avenae* was constructed for exploring more candidate parasitism genes. 5800 ESTs were generated and 2568 unigenes were obtained. Three β -1,4-endoglucanase genes (*Ha-eng-1a*, *Ha-eng-2* and *Ha-eng-3*) expressed in the pharyngeal glands of the sedentary cyst nematode (*H. avenae*) were cloned. The cDNA of *Ha-eng-1a* encoded a deduced 463-amino acid sequence containing a catalytic domain and a cellulose binding module separated by a linker. The genomic DNA of *Ha-eng-1a* is 2,129-bp long, containing eight introns ranging from 56 bp to 157 bp and nine exons ranging from 70 bp to 299 bp. Southern blot analysis revealed that two copies of the *Ha-eng-1a* gene are present in *H. avenae*. *In situ* hybridization showed that the *Ha-eng-1a* transcripts specifically accumulated in the two subventral gland cells of the second-stage juveniles.

Acknowledgement: This Research was supported by Chinese Special R & D Fund for Public Benefit Agriculture (200903040) and Ministry of Science and Technology (2009DFB30230).

4 Cereal cyst nematodes and its research perspectives in Iran

Tanha Maafi, Z¹, Ahmadi, A.² and Dababat, A.³

¹Iranian Research Institute of Plant Protection, P.O. Box 1454, 19395 Tehran, Iran .tanhamaafi@yahoo.com,

²Agricultural Research and Natural Resources Centre of Khuzestan, Ahvaz, Iran. alirahmadi2000@gmail.com;

³CIMMYT (International Maize and Wheat Improvement Centre), ICARDA CIMMYT Wheat Improvement Program, PO Box 39 Emek 06511 Ankara, Turkey. a.dababat@cgiar.org

Wheat is considered one of the most important strategic crops in Iran. Reducing the biotic and abiotic factors affecting the wheat yield production is one of the main priorities for the plant protection research sector in the country. Cereal nematodes are plant pathogenic agents combined with other fungal, viral and bacterial pathogens could cause severe damage on wheat production in Iran. Cereal cyst nematodes (CCN) have been identified in the main wheat producing regions, comprising of three species, in order of their abundances are *Heterodera filipjevi*, *H. latipons* and *H. avenae* type B. Progress has been made in the morphological and molecular identification, due to high overlapping in morphometric characters of *H. avenae* type A, B and *H. filipjevi* confirmation of morphological identification by molecular marker such as rRNA-ITS-RFLP has become an essential component of the recognition. Several research studies on the distribution and population density indicated that cereal cyst nematodes are widely distributed in most wheat producing fields with various population densities. The economic importance assessments demonstrate that these three species possess the potential to damage the grain wheat particularly under the rain-fed and drought conditions, the situations which were observed in most of the sampled regions. Although the damage threshold should be studied for each region and determined under different environmental conditions such as: availability of water and nutrition, genotypical factors *i.e* tolerance and resistance of cultivars. The other main priority in CCNs researches should be concentrated on the sources of genetic resistance to cereal cyst nematodes in Iranian cultivars and landraces of wheat that are unknown. Meanwhile, pathotype characterization of the Iranian populations of cereal cyst nematodes that could assist to some extent to determinant their genetic diversity is the other principal subject for achieving the resistance resources.

SESSION II The economic importance and biology of cereal nematodes

Chairman: Dr. Amer DABABAT, Prof. Jinling LIAO

5 The biology of Cereal Cyst Nematode in Hebei: implications on its control

Li Xiuhua Ma Juan Gao Bo Wang Rongyan Chen Shulong

Institute of Plant Protection, Hebei Academy of Agricultural and Forestry Sciences/ IPM Center of Hebei Province/ Key Laboratory of Integrated Pest Management on Crops in Northern Region of North China, Ministry of Agriculture, P. R. China, Baoding 071000, Hebei Province, China

Cereal cyst nematode is one of most important diseases in wheat production in Hebei, China. The biology of the nematodes was studied systematically for optimizing the control strategy. The nematode species were identified by morphological and molecular characteristics. The natural weed hosts in wheat field were investigated. The resistance of 41 local cultivars to the nematodes was evaluated in the pot and the field by counting the number of cysts produced. The hatch characteristics were tested with the cysts sampled from the natural fields in different periods. Effects of different temperatures on the nematode penetration and development were tested in mini pots. Life cycle was investigated in three fields for three years. The toxicity of different nematicides to eggs and second stage juveniles were tested in vitro. Effect of rotation, fallow and continues cropping on population dynamics of the nematodes were evaluated in natural fields. The results showed that only *Heterodera avenae* were detected in Hebei. *Avena fatua* and *Aegilops squarrosa* were the most important wild hosts for *H. avenae* in wheat fields. Three cultivars were resistant among the 41 cultivars tested, and four cultivars were highly tolerance to the nematode infection. The nematodes hatched at 5~10°C but not for over 15°C in 19 weeks for the newly formed cysts (June), while the hatching peak of *H.avenae* collected in wheat sowing period (October) was in 10~12 weeks and in 6~8 weeks at 5°C and 15°C respectively, but the optimum temperature for hatching of the nematode collect from natural field in December was 20°C. The optimum temperature for penetration is 16°C and the optimum temperature for nematode development and cyst forming is 18~22°C. Second stage juveniles could be extracted from soil all year round except the early and mid of June. There was a small peak of hatch before soil freezing, but most of nematodes hatched in the early and mid of April. The peaks of third stage juveniles, fourth stage juvenile and female occurred in the early, mid to late and late of May respectively. Second stage juveniles were more sensitive to the nematicides tested than the eggs or cysts. Population of *H. avenae* reduced by 89.8% after one year's fallow. The reducing rate was 93.8% after rotation with eggplant for one year, while it was 90.7% and 98.8% after rotation with the muskmelon or melon for one and two years respectively. In the natural disease fields, the population of the nematodes was increased by 36.8% and by 49.2% after wheat continuous cropping for one and two years respectively. The control of the *H. avenae* should integrate the resistance cultivars, weed eradication, fallow or rotation and chemical control.

Acknowledgments: We appreciate the financial support provided by the Ministry of Agriculture of China (200903040) and Ministry of Science and Technology of China (2009DFB30230).

6 Crop loss assessment of *Heterodera filipjevi* on some cultivars of wheat, barley and triticale under field condition of Southwest of Iran

Ahmadi, A.R.¹, Tanha Maafti, Z.², Dababat, A.³

¹Plant protection department, Khuzestan Agricultural and Natural Resources Research Centre, Ahvaz, Iran. alirahmadi2000@gmail.com;

²Iranian Research Institute of Plant Protection, Tehran, Iran. tanhamaafi@yahoo.com;

³CIMMYT (International Maize and Wheat Improvement Centre), ICARDACIMMYTWheat Improvement Program, PO Box 39 Emek 06511 Ankara, Turkey. a.dababat@cgiar.org

Khuzestan province is ranked as the first cereal producing province in Iran during 2011-2012. The cereal cyst nematode, *Heterodera filipjevi* is widespread in the main producing cereal fields in the region. It was found in 38% of the surveyed wheat fields of the province according to the recent studies. The damage assessment of *H. filipjevi* was performed on different yield components of four spring wheat cvs. Chamran, Virinak (bread wheat), Yavarus and Behrang (durum wheat), barley line wb/19/10 and Triticale cv. Juvanilo in an experiment under *Heterodera filipjevi* infested field in Ramshir region during 2008-2009. The trial was arranged in a randomized complete block design. Each treatment replicated seven times, consisting of plots with or without nematicide application (aldicarb 10G) at sowing time. The population density was determined before sowing to evaluate the initial population (Pi) and after harvesting for the final population (Pf). The obtained results indicated significant reduction of grain yield, biomass, shoot weight, shoot height and tillering by 52 (40-73), 40 (14-53), 38 (6-67), 15 (8-21) and 24 (10-39) percent in the different cultivars, respectively. The initial population was 10 eggs and juveniles/ g of soil. The nematode reproduction factor in plots plus and minus nematicide were ranged between 0.7-2.14 and 0.9- 4.52 respectively. This is the first evidence of the cereal yield losses caused by *H. filipjevi* in natural infested field conditions in Iran.

7 Importance of identification cereal cyst nematodes species and pathotypes in breeding for resistance and for management strategies

Ricardo Holgado¹ and Christer Magnusson²

¹Bioforsk, Norwegian Institute for Agricultural and Environmental Research, Plant Health and Plant Protection Division, Ås- Norway, ricardo.holado@bioforsk.no

²Bioforsk, Norwegian Institute for Agricultural and Environmental Research, Plant Health and Plant Protection Division Ås- Norway, christer.magnusson@bioforsk.no

Cyst nematodes are usually recognised by the morphology of the adult female (cyst) and by host plant associations. But, there are risks of misidentification if identification only rely on host plant. For more than 90 year the potato cyst nematode, the beet cyst nematode and cereal cyst nematode were considered as one species. In 1923 Wollenweber classified *Heterodera rostochiensis* as separate species. The cereal cyst nematode and the beet cyst nematode were considered as two races until 1940 when Franklin demonstrated a difference in length of second stage juveniles (J2). Later it was agreed to designate cereal cyst nematode as *Heterodera avenae* Wollenweber. It is known that cereals crops can be parasitised by a group of cereal cyst nematode involving a number of species, which differ morphologically and biologically. The most common and important species is *H. avenae*, but the “*Heterodera avenae*-complex” (Stone and Hill, 1982) includes also *H. arenaria*, *H. avenae*, *H. aucklandica*, *H. australis*, *H. bifenestra*, *H. filipjevi*, *H. hordecalis*, *H. latipons*, *H. mani*, *H. pratensis*, *H. spinicauda* and *H. ustinovi* (synonym *H. iri*). The virulence on cereal cultivars differs between and within species and several pathotypes occur among them. Borderlines between species and pathotypes often appear unclear. Identification usually requires morphometric studies including cyst size, colour, cuticular patterning, and several features of the cyst cone. The latter include the length of the vulval opening, the structure of the cyst wall around the vulva and in the perineal area, as well as internal structures of the vulval cone (bullae, underbridge, and vaginal sheath). Specific diagnostics also rely upon features of the J2, notably body length, number of lateral lines on the cuticle, and length and shape of the stylet and tail. Identification could be assisted by molecular approaches that can distinguish some species. Recognition and definition of pathotypes as a sub-specific grouping depend upon tests with plants with genetically distinct resistance named host differentials. True pathotypes are not yet distinguishable by molecular tools. Species of cereal cyst nematodes have had a long co-evolution with the host and progenitors. Consequently, there are many host resistance genes matching the virulence genes in the nematode populations. As breeding new cereals varieties may reduce the numbers of resistance genes; a decisive consideration for resistance breeding is to be aware of the degree of heterogeneity in cereal cyst nematode species and populations. Furthermore it is essential to pay attention to the fact that field population may contain several species and pathotypes.

Nematode management practices must be based on the knowledge of the population dynamics, the population density required to cause economic damage and the measures capable of reducing or keeping the population density below the threshold for economic damage.

In Norway management systems based on precise identification of nematode species and pathotypes and good knowledge on appropriate resistant cultivars are in operation. Resistant barley is generally recommended when nematode populations are high due to its high tolerance compared to resistant oats. Farmers implementing this program have reported increased cereal yields on the average of 1000 kg /ha. It has been calculated that by implementing this program in full, the county of Vestfold could make an economic gain of 800 000 € annually.

8 The effect of the cereal cyst nematode, *Heterodera avenae*, on wheat yield in Shangdong province

Zhang Jie Li Suxia Ren Yupeng Liu Feng*

College of Plant Protection, Shandong Agricultural University, Key Laboratory Pesticide Toxicology & Application Technique, Tai'an 271018, Shandong Province, China

Forevaluating the impact of *H. avenae* on yield of wheat in Shandong province of China, pot and field trials were conducted with different initial population densities (P_i) inoculated in soils. In pot trials, as the increasing of eggs per gram soil from 5-30, the chlorophyll contents of flag leaf decreased from 10.89% to 34.46%, dry weight of roots decreased from 30.65% to 73.95%, dry weight of shoots decreased from 13.93% to 58.68%, number of spikes decreased from 30.48% to 53.33%, grain numbers decreased from 3.97% to 36.93%, weight of spikes from 30.14% to 68.84%. The reproduction factors significantly decreased from 3.40 to 1.52 with the increase of initial egg density of cereal cyst nematode from 5 to 30 per gram soils, which negatively correlated with the production indexes. While under the field conditions in Feicheng city, there had no significant effect of different initial egg densities of cereal cyst nematode on the above production indexes of wheat.

Key words: Cereal cyst nematode; initial densities; yield, Fitting model

Acknowledgement: This Research was supported by Chinese Special R & D Fund for Public Benefit Agriculture (200903040).

9 Studies on cereal cyst nematodes, *Heterodera avenae* Wollenweber, in South East Anatolia and Eastern Mediterranean Regions in Turkey

İMREN M.¹, TOKTAY H.², DABABAT A.A.³, ELEKCIOĞLU I. H.⁴

¹ Biological Control Research Station, Yüreğir-Adana-TURKEY

² Faculty of Agricultural Sciences and Technologies, Nigde University, Nigde, Turkey,

³ Faculty of Agriculture, Department of Plant Protection Cukurova University, Balcalı Adana, Turkey,

⁴ CIMMYT (International Maize and Wheat Improvement Centre), Ankara, Turkey

Cereal cyst nematodes (*Heterodera* spp.) are a major constraint to cereal production in many parts of the world and contain at least 12 species that invade roots of cereals and grasses. *Heterodera avenae* (Wollenweber, 1924), *H. filipjevi* (Madzhidov, 1981), *H. latipons* (Franklin, 1969) are recognized as the most economically important species in this genera. *H. avenae* has wide distribution in temperate wheat-producing regions and it was first detected in Turkey in 1974 and is now known to occur in many cereal growing parts in Turkey. The objectives of this study were to focus on identification of cereal cyst nematodes (CCN) in cereal fields of South East Anatolian and Eastern Mediterranean region and finding out inter and intra species genetic variations of *Heterodera* species; to determine the biological features (hatching) of *Heterodera avenae* under in vitro; to investigate the pathotypes of *Heterodera avenae* using international host differential test materials in Eastern Mediterranean region; and to screen some wheat varieties (international, national wheat and wild wheat relatives) against *Heterodera avenae* and estimating yield loss caused by *H. avenae* in field condition. The result, based on phylogenetic analysis using ITS-rDNA sequences and morphological analysis; *H. avenae*, *H. filipjevi*, *H. latipons* species were identified. The highest cumulative hatching of 85.6% was obtained at a constant temperature of 15 °C for 283 days, the lowest cumulative hatching of 16.6% was obtained at 25 °C for 283 days. Nematode populations were found completely belong to Ha1 group Ha21 pathotype and this study is the first report to determine *H. avenae* pathotype in Turkey. Four national wheat varieties, seventeen wheat wild genotypes and twenty three international wheat genotypes were found to be moderately resistant against Ha1 group, Ha21 population of *H. avenae*. The average yield loss varied from 1,19% to 21,8% in Eastern Mediterranean region.

Key words: *Heterodera* spp., hatching, pathotype, screening, yield loss

10 Spatial distribution of the cereal cyst nematodes in Egyptian bread wheat (*Triticum aestivum* cv. Giza 68) field

M.M.M. Abd-Elgawad and M.M.M. Mohamed

Phytopathology Department, National Research Center, El-Tahrir St., Dokki 12622, Giza, Egypt

Corresponding author: mahfouzian2000@yahoo.com

The spatial distribution of the cereal cyst nematodes, *Heterodera* spp., in 12 different 350-m²-plots of bread wheat (*Triticum aestivum* cv. Giza 68) field was studied. The dispersion parameter, viz. mean–variance test, of the nematode larvae in soil indicated agreement with a Poisson model at the 95% probability level and the hypothesis of randomness was not disproved in any of the sampled plots. Chi squared (χ^2) test for goodness-of-fit across all the plots proved that the same model was a good fit to the original nematode counts too. Yet, Taylor's Power Law was not fit ($P \leq 0.05$) to *Heterodera* spp. population data obtained from the plots. Sample size optimization needed to achieve a predetermined level of sampling error for the nematodes was calculated. For example, to sample cereal cyst nematode larvae with a 0.20 or 0.1 standard error to mean ratio and 11 nematodes/100 gm soil, one could collect 2 or 7 samples of 5 cores each, respectively. Integrated nematode management practices in the light of their random distribution were discussed.

11 Field dynamics and hatching characteristics of two speices of wheat cereal cyst nematode in Henan province

H.-T. Yan, H.-L.Li, H.-X.Yuan, X.-P.Xing and B.-J. Sun

Department of Plant Pathology, Henan Agricultural university, NO.95 Wenhua Road, Jinshui District, Zhengzhou, 450002, P. R. China

The distribution and infection dynamics of *Heterodera avenae* and *H. filipjevi* were studied in the fields in Henan province. The hatching characteristics of *H. avenae* and *H. filipjevi* at different temperatures were studied. The preliminary results were as follows.

(1) The test field in Xuchang(*H. filipjevi*) had 60 plots, the field in Xingyang(*H. avenae*) had 48 plots, and each plot was 49m². The horizontal samples analysis result indicated that every plot had cereal cyst nematodes, but the densities of cyst were different in different plot. The highest cyst density was 36 per 100g soil, the lowest was 4.3 per 100g soil in Xuchang field. The highest cyst density of plot was 38.7 per 100g soil, and the lowest density of plot was 4.3 per 100g soil in Xingyang field. The vertital samples analysis result indicated the number of cysts was the highest in 5-15cm soil layer. The number of cysts was tapering along with depth increasing of soil layer. The cysts of field in Xingyang were mainly distributed in 10-15 cm soil layer, the average number was 98.3 per 100g soil. The cysts of field in Xuchang were mainly distributed in 5-10 cm soil layer, the average number was 66.7 per 100g soil.

(2) The wheat roots were infected by the second-stage juveniles two weeks (early November) after planting in the two fields. A few third-stage juveniles were found four weeks after planting. The first infection peak of the second-stage juveniles comes in the 6th week after the planting. With the temperature got lower 60 days after planting in the winter, the number of the larva of different stages remained stable. 120 days after planting, with the temperature rising in the spring, the number of the second stage juveniles started to increase and reached to the second peak in the late March and early April, but the number of juveniles which invaded the wheat roots in the second peak were apparently less than the first one. The larva gradually developed into white females and cysts. The infection dynamics of two species was approximately alike, but the third stage, fourth stage of the larva and the white females of the *H. filipjevi* appeared one week earlier than that of *H. avenae*.

(3) The cysts were collected from the fields of Xuchang and Xingyang in the middle of August. The influence of the temperature on the hatching of the two species was studied indoors. The results showed that *H. avenae* could hatch at 5°C, 10°C, 15°C, 20°C and 25°C, *H. filipjevi* could hatch at 5°C, 10°C and 15°C, but it could hardly hatch at 20°C and 25°C. The most suitable temperature of hatching was at 10°C or 15°C. The second-stage juveniles started to emerge on the 20th day. The hatching peak appeared on the 50th to the 60th day. There were more second-stage juveniles of *H. filipjevi* than that of *H. avenae*. The hatching continued until the 75th day. Changing the temperature could increase the number of the second-stage juveniles dramatically, and the hatching rate reached 83%.

Acknowledgement: This Research was supported by Chinese Special R & D Fund for Public Benefit Agriculture (200903040).

12 The effect of the rotation with peanut on CCN population and wheat yield

Honghai ZHAO¹, Haiyan DING¹, Chen LIANG¹, Deliang PENG²

¹College of Agronomy and Plant Protection, Qingdao Agricultural University, Qingdao 266109, China;

²State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, CAAS, Beijing, 100193, China

Heterodera avenae (CCN) is the most important nematode damaging wheat in China. It was numerously reported that juvenile emergence from eggs stopped in newly formed brown cysts and the break of dormancy required low temperature treatment for a relative long time. Free juveniles were also found out of the egg shell in newly formed brown cysts with different juvenile-egg ratio during our CCN investigations. The test was implemented to determine the activity of free juveniles and eggs in newly formed brown cysts. Free juveniles and eggs were isolated under room temperature (22~30 °C), afterwards some of them were inoculated onto wheat seedlings immediately and some were treated at low temperature (5°C) for different length of time before the inoculation. Wheat seedlings were earlier planted in paper cups, and incubated under 15~16°C in light incubator after each inoculation. Wheat roots were taken out and stained to detect infection situations after 3~4 weeks of incubation. It was found that no infection occurred for both juveniles and eggs without low temperature treatment; infection occurred only for the juveniles which had been treated with low temperature for 20, 30 and 45 d, and for the eggs treated with low temperature for 10, 20, 30 and 45 d. The results indicated that dormancy happened on the juveniles as well as eggs in newly formed brown cysts and the encysted juveniles required low temperature treatment to activate the ability of infection or even emergence from cysts.

Key words: *Heterodera avenae*; newly formed brown cysts; encysted juvenile; dormancy

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

13 Morphometrical and molecular characterization of root lesion nematodes (*Pratylenchus* spp.) on wheat in Morocco

F. Mokrini^{1,2}, L. Waeyenberge², N. Viaene^{2,4}, F. Abbad Andaloussi⁵ and M. Moens^{2,3}

¹National Institute of Agricultural Research, (INRA), Km 9, 14000 Kenitra, Morocco.

² Institute for Agricultural and Fisheries Research, Plant, Crop Protection, Burg. Van Gansberghelaan 96, B-9820 Merelbeke, Belgium.

³ Laboratory for Agrozoology, Ghent University, Coupure links 653, B-9000 Ghent, Belgium.

⁴Department of Biology, Ghent University, K.L. Ledeganckstraat 35, B-9000 Ghent, Belgium

⁵National Institute of Agricultural Research, Scientific Division, BP415RP, Rabat Morocco

Corresponding author: fmokrini@yahoo.com

Root-lesion nematodes of the genus *Pratylenchus* have a worldwide distribution and are regarded as severe production constraints for numerous important crops. A rapid and accurate method to identify *Pratylenchus* to the species level is necessary to develop appropriate management strategies. During a survey of wheat-growing areas of Morocco from May to June 2011, 18 populations of the root-lesion nematode were collected. The nematodes were extracted from root and soil using an automated apparatus based on centrifugal floatation. They were identified on the basis of their morphological and morphometrical characters, complemented by molecular methods. The morphometrical observations of the collected females and males demonstrated the occurrence of *Pratylenchus penetrans* in most of the samples; *Pratylenchus thornei* and *P. pseudo coffeae* were only detected in samples from Zaere and Settat, respectively. After morphometrical identification, DNA was extracted from a single individual that was hand-picked. The duplex PCR primers described by Waeyenberge *et al.* (2009) were used to identify *P. penetrans*; the species-specific forward primer PTHO and the common reverse primer D3B (Al-Banna *et al.*, 2004) were used to identify *P. thornei*. For the remaining populations that were not identified by species-specific primers, the D2D3 expansion segments of the 28S rRNA gene were amplified with the forward D2A and reverse D3B primers (Joyce *et al.*, 1994). The purified PCR products were sequenced using the same primers. The obtained sequences were compared with those of *Pratylenchus* species available in the GenBank database (www.ncbi.nlm.nih.gov). This comparison confirmed the morphological identifications. The study of the phylogenetic relationship of the Moroccan *P. penetrans* populations together with other populations of *P. penetrans* and some species of which the sequence was available in GenBank (NCBI) showed a high similarity (98,5%) between all *P. Penetrans* populations from Morocco. This is the first report on molecular characterization of *Pratylenchus* populations from Morocco.

Session III: Control strategies of CCN in wheat using host resistance
Chairmen: Prof. Richard.W. SMILEY, Prof. Hongmei LI

14 Screening of International spring Wheat Germplasms against the Cereal Cyst Nematodes (*Heterodera avenae*) and the Root Lesion Nematodes (*Pratylenchus thornei* and *P. neglectus*) in-vitro Conditions in 2012, Turkey

ELEKCIOĞLU I. H.¹, İMREN M.², TOKTAY H.³, BOZBUĞA R.², BRAUN H. J.⁴, DABABAT A.A.⁴

¹ Faculty of Agriculture, Department of Plant Protection Cukurova University, Balcalı Adana, Turkey,

² Biological Control Research Station, Yüreğir-Adana-TURKEY

³ Faculty of Agricultural Sciences and Technologies Niğde University, Niğde, Turkey,

⁴ CIMMYT (International Maize and Wheat Improvement Centre), Ankara, Turkey

The occurrences and distribution of Cereal cyst nematodes (*H. avenae*, *H. filipjevi* and *H. latipons*) and Root lesion nematodes (*P. thornei* and *P. neglectus*) have been surveyed in research projects in different part of Turkey. The most environmentally sound methods to control these nematodes is the use of resistant germplasm. Therefore, international, varieties have been screened for their resistance reactions to those nematodes at the Biological Control Research Station in Adana-Turkey. The International Maize and Wheat Improvement Centre (CIMMYT), Turkish Food, Agriculture and Livestock Ministry and in collaboration with Cukurova University have been conducted screening project to understand the resistant reactions of International spring wheat varieties and lines against to *H. avenae*, *P. thornei* and *P. neglectus* in nematology part of the Soil Borne Pathogen Project since 2001. In this aspect 29 SAWSN was investigated resistance against to these nematodes in 2012. The experiments were established with 7 replicates and two biological repetitions. Each line were germinated and planted individually in small tubes filled with 80 g soil mixture. 400 nematodes for *P. thornei*, *P. neglectus* were inoculated to each tubes on seventh planting day and 200 J₂ for *H. avenae* onplanting day. Plants harvested after 9-12 weeks and the number of *P. thornei*, *P. neglectus* and *H. avenae* cysts and juveniles (J₂) per plant were counted. Totally 5292 seeds were tested against to *P. thornei*, *P. neglectus* and *H. avenae* during the 2012 screening program. The results indicated that several wheat lines with multiple resistances to *H. avenae*, *P. thornei* and *P. neglectus* have been identified. This result gave significant contribution to genetic resistance against *H. avenae*, *P. thornei* and *P. neglectus* in international wheat breeding programs.

Key words: Screening, Resistance, *H. avenae*, *P. thornei*, *P. neglectus*

Acknowledgment: CIMMYT and the Turkish Ministry of Agriculture would like to thank the Grain Research and Development Corporative GRDC for funding this project and Syngenta for partial money support to present this work at this meeting.

15 Toward management of cereal cyst nematode with host resistance: identification of effective resistant sources in China

Hongjie Li¹, Lei Cui¹, Honglian Li², Xiaoming Wang¹, Wenhua Tang³

¹ Institute of Crop Science, Chinese Academy of Agricultural Sciences, Beijing 100081, P. R. China

² College of Plant Protection, Henan Agricultural University, Zhenzhou 450002, P. R. China

³ Department of Plant Pathology, China Agricultural University, Beijing 100193, P. R. China

Corresponding author: lihongjie@caas.cn

Recent prevalence of cereal cyst nematode (CCN, *Heterodera avenae* and *H. filipjevi*) throughout the important wheat growing regions of China has attracted serious attentions. Unfortunately, the shortage of resistant resources has limited the use of host resistance to control CCN. Since 2008, a project has been initiated to identify effective sources of resistance to the prevalent pathotypes of *Heterodera* spp. indigenous to China. Based on a three-year field test, resistance of the wheat cultivar Madsen from Washington State University, USA was effective against both species of *Heterodera*. Results of histological analysis indicated that the number of juveniles invaded the roots of Madsen was fewer than the susceptible control Wenmai 19. Inoculation tests with 11 pathotypes of *H. filipjevi* and *H. avenae* from Henan, Anhui, and Shandong provinces demonstrated that Madsen was effective against all the populations of both species of *Heterodera*. Genetic analysis indicated that a single dominant gene was associated with the resistance of Madsen to *H. filipjevi*, which permits the transfer of its resistance to CCN in to more adapted local cultivars. Some advanced lines have been produced, which have shown to be resistant to *H. filipjevi* and *H. avenae* as effective as the donor parent Madsen. In addition to resistance to *H. filipjevi* in Madsen, a number of *Triticum durum* and wheat-*Thinopyrum* derivatives, *Triticale*, *T. dicoccum*, and *T. dicoccoides* accessions exhibited excellent resistance against CCN in the field tests. Since most commercial wheat cultivars currently grown were susceptible, these resistant germplasms provide valuable sources for improvement of CCN resistance in China.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

16 New sources of Resistance against the Root Lesion Nematodes, *Pratylenchus thornei* Sher et Allen in some National and International Wheat Germplasm

TOKTAY, H.¹, IMREN, M.², BOZBUĞA, R.², DABABAT, A.³, ELEKCIOĞLU, I. H.⁴

¹ University of Niğde, Faculty of Agricultural Sciences and Technologies, Niğde, Turkey,

² Biological Control Research Station, Yüreğir-Adana-TURKEY

³ CIMMYT (International Maize and Wheat Improvement Centre), Ankara, Turkey

⁴ Faculty of Agriculture, Department of Plant Protection Cukurova University, Balcalı Adana, Turkey.

Root Lesion Nematodes (RLNs) are widespread throughout the grain growing regions of Turkey. *Pratylenchus thornei* Sher et Allen is the predominant species of RLN in the and causes a yield losses of up to 19% in the Eastern Mediterranean Region. Due to the huge range of hosts to the RLN makes crop rotation is a weak option to control its damage. Therefore, breeding wheat germplasms for resistance to this destructive nematode will be an effective and economical method of minimizing crop losses by preventing nematode reproduction and reducing the overall disease problem by leaving fewer nematodes in soils to attack subsequent crops. Globally there are several known sources of genetic resistance in wheat against root lesion nematodes, However, these sources are still not well documented whether they are effective in Turkey or not. In this study, 90 international and 30 national wheat varieties have been screened under the *in vitro* conditions against a common root lesion nematode (*P. thornei*) at the Biological Control Research Station in Adana-Turkey. The results of this screening indicated that eight national wheat varieties and fifty-nine international wheat genotypes were found to be moderately resistant against Adana population of *P. thornei*. Among these genotypes, the national bread wheat variety, Adana 99 (PFAU/SERI82//BOG"S") approved gave resistant reaction to *P. thornei*. In conclusion, those national and international genotypes are of high value to the Turkish and regional breeding programs. However, testing more varieties against more population of *P. thornei* is still needed.

Key words: Root Lesion Nematodes, *Pratylenchus thornei*, Screening.

17 Identifying a novel source of resistance against *Heterodera filipjevi* in spring barley ‘Steptoe’ by analysis of a double haploid population Steptoe × Morex

A. Westphal¹ and A. Graner²

¹Institute for Plant Protection and Grassland, Julius Kühn-Institut, Messeweg 11/12, 38104 Braunschweig, Germany

²Leibniz Institute of Plant Genetics and Crop Plant Research (IPK) Corrensstrasse 3, 06466 Gatersleben, Germany

Corresponding author: andreas.westphal@jki.bund.de

Cereal cyst nematodes (CCN) damage small grain crops in many regions of the world. In Europe, these pests have become known for their infectivity on emerging spring cereals. A newly recognized species of the CNN complex is *Heterodera filipjevi*, which is similarly spread as *H. avenae* and causes damage in arid areas among others in the Pacific North West of the US, the Mediterranean and China. Resistance to *H. avenae* may not be effective against *H. filipjevi*. A resistant response to *H. filipjevi* was found in *Hordeum vulgare* ‘Steptoe’ while this cultivar was susceptible to *H. avenae*. The objective of the current study was to determine the chromosomal location of the resistance to *H. filipjevi* in Steptoe by use of a Steptoe × Morex double haploid population. In the greenhouse in small growth containers, a total of 93 double haploid lines, along with the resistant and susceptible parents, were tested if they permitted *H. filipjevi* to reproduce. Genetic mapping using a dense framework of single nucleotide polymorphism (SNP) markers allowed for precise positioning of the resistance locus on chromosome 3H. The development of markers appropriate for marker assisted selection will help breeders to effectively incorporate resistance into elite germplasm.

18 Selection of Certain Wheat Genotypes for Resistance to Cereal Cyst Nematode (*Heterodera avenae*) Based on growth parameters and Molecular Markers

Moustafa, Kh.A.^{1,3}, Al-Doss, A.A.^{1,3}, Motawei, M.I.², Al-Otayk, S.², Dawabah, A.A.M.^{2,3*}, Abdel-Mawgood, A.L.^{1,3}, Al-Rehiyani, S.M.² and Al- Hazmi , A.S.^{2,3}

¹Plant Production Department, College of Food and Agriculture Sciences, King Saud University, Saudi Arabia,

²Plant Production and Protection Department, College of Agriculture and Veterinary Medicine, Qassim University, Saudi Arabia,

³Centre of Excellence in Biotechnology Research, College of Food and Agriculture Sciences, King Saud University, Saudi Arabia

Corresponding author: adawabah@ksu.edu.sa

The cereal cyst nematode (CCN), *Heterodera avenae* Woll. causes severe damage to wheat (*Triticum aestivum* L.) production in the Kingdom of Saudi Arabia. This study aimed to screen certain wheat genotypes for resistance to CCN to aid in the development of new resistant cultivars of bread wheat adapted to Saudi Arabian conditions. The field performance of 17 genetically diverse wheat genotypes (local and international materials) were evaluated for two successive years (2009 and 2010) in a *H. avenae*-naturally-infested field. Results showed that the tested wheat genotypes were significantly different in their field performance, and in their resistance to CCN. Ten local wheat genotypes were designated as resistant. The local cv. KSU 119 was the most resistant one (no. cysts/plant= 0.7) among all the tested genotypes. However, the Australian cv. AUS-30851 and the SIMMYT cvs. 15 SAWYT-30, 15 SAWYT-31, 15 SAWYT-38, and 15 SAWYT-42 plus the susceptible standard Yecora Rojo were found to be the most susceptible (no. cysts/plant= 18-28) genotypes in this study.

Microsatellite markers linked to *Cre1* and *Cre3* genes were used in this study. It was found that ten out of 17 wheat genotypes (LNM-72, LNM-99, LNM-126, LNM-136, KSU118, L11-8, L11-17, L11-21, KSU 119, and AUS-30851) had both *Cre* genes. The dendrogram generated using SSR data divided wheat genotypes into two main clusters. Genotypes LNM-72, LNM-99, LNM-126, LNM-136, KSU118, L11-8, L11-17, L11-21, and KSU 119 were found in the same sub-cluster. These genotypes were found to be the most resistant to CCN. Therefore, amplification conditions for *Cre3* and *Cre1* loci were optimized and are now used in our marker-assisted selection (MAS) programs to identify CCN-resistant wheat genotypes.

Key words: Wheat, *Heterodera avenae*, Pathotypes, PCR, Marker Assisted Selection (MAS).

Session IV: Control strategies of CCN other than host resistance
Chairmen: Dr. Ricardo HOLGADO, Dr. Yuxi DUAN

19 Influence of abamectin and *Bacillus firmus* on *Heterodera avenae* and on spring wheat yields in the USA

R.W. Smiley, J.M. Marshall and G.P. Yan

Oregon State University, Pendleton, Oregon, 97801, U.S.A.

Email: richard.smiley@oregonstate.edu

The cereal cyst nematode, *Heterodera avenae*, reduces wheat yields in the Pacific Northwest (PNW) states of Idaho, Oregon and Washington. Documentable annual economic losses have been estimated at US\$3.4 million but actual losses are thought to be much greater. Seed treatments that are effective against plant-parasitic nematodes on other crops were evaluated on spring wheat. Abamectin (Avicta) and *Bacillus firmus* (Votivo) were examined in field trials in each of the PNW states. Aldicarb (Temik), banded with the seed, was included as a treatment for comparative purposes. The initial density of *H. avenae* eggs plus juveniles, from cysts, in the three fields averaged 2332, 2860, and 6666/kg of soil. The first two locations were also infested by high, economically important densities of *Pratylenchus neglectus*; 2602 and 13596/kg of soil, respectively. Abamectin and *B. firmus* had no effect on grain yield or on the incidence or severity of the root knotting symptom caused by *H. avenae*. These products also failed to reduce the post-harvest density of *H. avenae* or *P. neglectus*. Aldicarb increased the mean grain yield by 949 kg/ha, valued at \$312/ha. Aldicarb also reduced the severity of root symptoms and reduced the post-harvest density of *H. avenae* by 38% to 93%. However, aldicarb is not, and cannot, be registered for commercial use on wheat in the USA. We concluded that management of cereal cyst nematode can be achieved most effectively through further emphasis on crop rotations and on development of wheat cultivars with both resistance and tolerance to *H. avenae*.

20 Effect of essential oils of some medicinal plants on phytonematodes in Egypt

M.M.M. Abd-Elgawad¹, E.A. Omer² and R.F. Ismail²

National Research Centre, Dept. of Plant Pathology¹, Pharmaceutical Sciences Dept. ², Dokki 12622, Giza, Egypt.

Corresponding author: mahfouzian2000@yahoo.com

The phyto-extracts have advantages over synthetic nematicides because of their new compounds – with usually less concentrate than synthetic chemicals - that nematodes are not able to inactivate, multiple action modes, and formation of renewable sources. Therefore, ethanol extract of *Tagetes lucida*, a medicinal plant recently introduced into Egypt, was tested and compared with other plant species *Achillea millefolium*, *Cymbopogon citratus*, *Artemisia annua* and *Calendula officinalis* for their nematicidal activity against plant-parasitic nematodes of common occurrence. All extracts inhibited ($P \leq 0.05$) motility of *Meloidogyne incognita*, *Criconebella* spp., *Helicotylenchus* spp., and *Pratylenchus* spp. *A. annua* was generally more effective in reducing the numbers of active nematodes tested than others except *T. lucida*-root extract which was superior to herbal extract. When transferred to water, the total nematodes that regained their activeness ranged from 15% for *T. lucida* to 38% for *A. millefolium* after 24 hours and from 4% for *C. officinalis* to 39% for *A. millefolium* after 72 hours. The extracts inhibited *M. incognita*-juvenile hatching in the range 46.8% for *A. millefolium* to 88.8% for *C. citratus* compared to 5.1% at the controls. The nematicidal activity of the isolated materials from the plants was concentration dependent. The main compound(s) of each plant extract determined by GLC analysis was presented and their mechanisms of action were discussed. Further studies are warranted to obtain insights on rates and timing of their possible application as bio-nematicides, as well as growth parameters of treated plants under greenhouse then field conditions in Egypt.

21 *Verticillium chlamydosporium*, a fungal parasite of the Cereal Cyst Nematode (*Heterodera avenae*) in the Saudi Fields

Al-Hazmi, A.S.¹, Dawabah, A.A.M.² and Al-Nadary, S.N.^{3*}

¹Ahmad Sa'ad Al-Hazmi: Plant Protection Department, College of Food and Agriculture Sciences, King Saud University, Saudi Arabia, asalhazmi@ksu.edu.sa

²Ahmed Abdel-Samie Mohamed Dawabah: Plant Protection Department, College of Food and Agriculture Sciences, King Saud University, Saudi Arabia, adawabah@ksu.edu.sa

³Saleh Noaman Al-Nadary: Plant Protection Department, College of Food and Agriculture Sciences, King Saud University, Saudi Arabia, nadary44@gmail.com.

Corresponding author: adawabah@ksu.edu.sa

Cereal cyst nematode (*Heterodera avenae*) is a devastating root parasite of wheat and barley in Saudi Arabia. During a survey of *H. avenae* in the infested wheat and barley fields in Hail and Tabuk regions, north Saudi Arabia, we noticed a heavy colonization of some extracted cysts with a fungal mycelium. The fungus colonized the encysted eggs as well, and was morphologically identified as *Verticillium chlamydosporium*. The fungus had optimum growth on yeast extract peptone-glucose liquid medium at 25 °C. Pathogenicity of the fungus on the eggs and newly formed cysts of *H. avenae* was evaluated under laboratory conditions. Further studies are being carried-out to identify the strain of the fungus on molecular basis, and also to determine the role of the fungus as a biocontrol agent for *H. avenae* on wheat plants

Key words: barley, egg parasites, infected cysts, nematophagous fungi, wheat

22 Toward Integrate management of cereal cyst nematode (CCN) in China: an example for research work in Jiangsu Province

H.-M. Li, X. Wang, X.-D. Liang and Y.-K. Chi

Department of Plant Pathology, Nanjing Agricultural University, Nanjing 210095, China

Corresponding author: lihm@njau.edu.cn

The cereal cyst nematode on wheat was first discovered in Jiangsu Province during May of 2009. The wheat growing area of Jiangsu province is about 2.3 Mha and listed as 5th in China. The well understanding of disease caused by CCN is urgently needed for making effective control strategies.

A systematic investigation of CCN distribution was carried out during 2009-2011. CCN was detected from 301 out of 580 investigated fields which from 152 towns of 47 counties in Jiangsu province. The widely distribution has caused serious damage to wheat production and the population densities of CCN in some areas were much higher than the economic threshold of losses. All Jiangsu populations of CCN were identified as *Heterodera avenae* by morphological and molecular characters and the status of *H. filipjevi* is unknown.

The life cycle of *H. avenae* on winter wheat in Peixian, Jiangsu Province was investigated systematically during two wheat growing season in 2010-2012. Peixian population finished only one life cycle during the whole growth season. Only few hatched second stage juveniles (J2) infected roots before wheat over-wintering and none of them developed in normal. The hatch peak appearing in soil and massive infection to roots was happened at the end of February in each year. At attassel period, the white females appeared on roots which can be seen by naked eyes and some males can be observed from roots and soil. At maturation period of wheat, the white females changed into brown cysts and fell into soil for over-summering.

Planting resistant cultivars is the most economical and practical measure for controlling the soil-borne disease caused by CCN. Optimizing the conditions of bioassay is important for the accuracy of resistance/susceptible evaluation for wheat cultivars. The cysts pre-treated at 4°C for 8 weeks and further incubated at 15°C obtained a large amount hatching J2 in short period, which can be used as inoculum for bioassay. The wheat seedlings planted in different size of containers with inoculum density of 4 juveniles per cm³ soil produced the largest number of white females. The screening bioassay using the optimized conditions revealed only Huamai No.1, Wenliang 58 and Yumai 66-18 out of 40 wheat cultivars was evaluated as highly resistant to Peixian population of *H. avenae*. At the meantime, the field trial evaluated only Huamai No.1 as resistance, which can be used as the prospective resistant cultivar planting in CCN heavily infested area of Jiangsu province.

The control effect of five granular nematicides (GR) with different usages was evaluated on CCN in wheat field during regreening stage. Although the numbers of cysts in soil collected from different treatments were clearly dropped after nematicide application, there is no significant difference in corrected cyst reduced rates between different nematicide treatments. Due to the effect of 0.5% Avermectin GR on inhibiting the reproduction of nematodes and improving the growth of wheat, the usage of 30 kg/ hm² was suggested to be applied in regreening field heavily infested with CCN, which might decrease the yield loss to some extent. Furthermore, economical seeds-coating chemicals were screened and tested for CCN control. The result revealed the self-patented Gannong seed coating III not only has the better control efficacy for CCN, but also has characteristics of environmental safety, lower toxicity, labor and cost saving, which is suitable for widely application in China.

Key words: *Heterodera avenae*; distribution; species identification; life cycle; bioassay optimization; resistance evaluation; integrated control

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

Session V: Molecular diagnosis of cereal cyst nematodes
Chairmen: Prof. Andreas WESTPHAL, Prof. Heng JIAN

23 Sensitive and direct detection of *Heterodera filipjevi* in soil and infected wheat by species-specific SCAR-PCR Assays

Huan Peng¹, Xiaoli. Qi¹, Gaofeng Wang¹, Deliang Peng¹, Haibo Long², Xufeng He¹,

Wenting He¹

¹ State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China

² Key Laboratory of Pests Comprehensive Governance for Tropical crops, Environment and Plant Protection Institute, Chinese Academy of Tropical Agricultural Science, Danzhou 571737, China

Corresponding author: pengdeliang@caas.cn

Cereal cyst nematodes (CCN), especially *Heterodera avenae* and *H. filipjevi*, are the most economically important plant-parasitic nematode on cereal crops in wheat production area of the world. Morphological identification these species is time-consuming and laborious because there are only slight differences. In this study species-specific SCAR-PCR assay for detection and identification of *H. filipjevi* from infected wheat roots and soil were developed. The species-specific primers were designed according to the randomly amplified polymorphic DNA (RAPD) markers amplified with random primer OPK16. A 646bp specific fragment of sequence was generated, which characterized amplified regions (SCAR) in *H. filipjevi*. The detection limitation of PCR assay was as low as 0.125 µl second-stage juvenile lysate, 3.9×10⁻³µl adult female lysate and 10⁻³ µl cyst lysate. The method was able to detect the various developmental stages of *H. filipjevi*, and a single of nematode in 0.5g soil. Two of six field samples (TYHN and XCHN) were detected as *H. filipjevi* by the method. In addition, we determined that the TYHN sample contained a mixed population of *H. filipjevi* and *H. avenae*. This present study is the first to provide a definitive diagnostic assay for *H. filipjevi* in wheat roots and soil using SCAR primer. The discovery of *H. filipjevi* in the Tangyin, Anyang city, Henan province represents a new record for the occurrence of this species in China.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

24 Quantitative detection of *Heterodera avenae* and *H. filipjevi* using qPCR

F. Toumi^{1,2}, *L. Waeyenberge*^{1,2}, *N. Viaene*^{1,2}, *A. Dababat*³, *J.M. Nicol*³, *F.C. Ogbonnaya*⁴ and *M. Moens*

¹*Institute for Agricultural and Fisheries Research (ILVO), Burg. Van Gansberghelaan 96, 9820 Merelbeke, Belgium; fateh.toumi@ilvo.vlaanderen.be;*

²*Faculty of Sciences, Ghent University, K. L. Ledeganckstraat 35, 9000 Ghent, Belgium*

³*International Maize and Wheat Improvement Centre (CIMMYT), Ankara, Turkey; a.dababat@cgiar.org; j.nicol@cgiar.org*

⁴*International Center for Agricultural Research in the Dry Areas, PO Box 5466, Aleppo, Syria. *Current address: GRDC, Level 1, 40 Blackall Street, Barton, ACT 2600, Australia; francis.ogbonnaya@grdc.com.au*

Corresponding author: fateh.toumi@ilvo.vlaanderen.be

Twelve *Heterodera* species are considered of major economic significance in wheat and barley. *Heterodera avenae*, *H. filipjevi* and *H. latipons* are the most important ones. Precise identification and quantification of these nematodes are necessary to develop effective integrated pest control. We report the results of a qPCR assay that we developed for the quick detection and quantification of *H. avenae* and *H. filipjevi*. Two qPCR primer sets comprising two primers and a probe, were designed and optimized. Both developed qPCR were able to detect a single second stage juvenile (J2). Their specificity was confirmed by the lack of amplification of DNA extracted from 13 other *Heterodera* species. A qPCR using DNA extracted from 106 and 114 J2 + mature eggs of *H. avenae* and *H. filipjevi*, respectively, resulted in steady Ct-values ($Ct = 22.33 \pm 0.1$ and $Ct = 21.83 \pm 0.05$, respectively). Dilution series of DNA extracted from known numbers of J2 of *H. avenae* and *H. filipjevi* were made. The qPCR resulted in a standard curve showing a highly significant linearity between the Ct-values and the dilution rates ($R^2 = 0.99$; slope = -3.05 and $R^2 = 0.99$; slope = -3.4 for *H. avenae* and *H. filipjevi*, respectively). A final validation test showed a high correlation between real numbers of J2 in a sample and the numbers detected in that sample by qPCR. The two qPCR provide a sensitive tool for the rapid detection and quantification of both species whether they occur alone or in mixed populations with other *Heterodera* spp.

Keywords: Nematode, qPCR, *H. avenae*, *H. filipjevi*.

25 Developing species-specific PCR assays for identification of *Heterodera filipjevi* and *H. avenae*

Guiping Yan, R.W. Smiley, P.A. Okubara and A.M. Skantar
Oregon State University, Pendleton, Oregon, 97801, U.S.A
Corresponding author: richard.smiley@oregonstate.edu

Heterodera avenae is an economically important cyst nematode that restricts production of cereal crops in the Pacific Northwest USA. *H. filipjevi* also occurs in winter wheat fields in Oregon. Identification of these two species is important for recommending and implementing effective management practices. Primers were designed from the internal transcribed spacer (ITS) regions of *H. avenae* and *H. filipjevi* ribosomal DNA. The primers were specific when tested with DNA from forty isolates belonging to ten *Heterodera* spp., five *Globodera* spp., five *Meloidogyne* spp., four *Pratylenchus* spp., and three other plant-parasitic nematode species. *H. filipjevi*-primers were also predicted to be highly specific using *in silico* analysis and sixty ITS sequences of closely related *Heterodera* spp. from many other countries. PCR reaction and amplification conditions were established, and *H. avenae* and *H. filipjevi* were clearly distinguished by unique PCR amplicons specific to each target species. Robust PCR amplification was achieved with DNA extracted from a single egg or second-stage juvenile (J2) using a laboratory-made worm lysis buffer, and DNA from 0.5 egg or 0.5 J2 using a commercial kit. The PCR assays were successfully employed for the differentiation of *H. filipjevi* and *H. avenae* populations collected from eight locations in three Pacific Northwest states (Oregon, Washington, and Idaho). This is the first report of a species-specific PCR assay to detect and identify *H. filipjevi*. The species-specific end-point PCR assays for *H. filipjevi* and *H. avenae* will enhance the diagnosis of cereal cyst nematode species in infested fields.

Session V: Genome and parasitism genes
Chairmen: Prof. Mike JONES, Dr. Deliang PENG

26 Gene silencing in root lesion nematodes significantly reduces reproduction in host plants

M.G.K.Jones, J-A C.W. Tan and Fosu-Nyarko, J.

Plant Biotechnology Research Group, School of Veterinary and Life Sciences, Western Australian State Agricultural Biotechnology Centre, Murdoch University, Perth WA 6150, Australia

Corresponding author: m.jones@murdoch.edu.au

Root lesion nematodes (RLNs, *Pratylenchus* species) are major root pests of many crop plants such as wheat and sugarcane. In Australia *P. thornei*, *P. penetrans* and *P. neglectus* can reduce wheat yields by 7-15%, and *P. zae* reduces sugarcane yields by a similar amount in fine textured soils. To study the potential of applying gene silencing technology to RLNs, the transcriptome of *P. thornei* has been sequenced using Roche 454FLX technology and annotated (Nicol, P. et al 2012, *Int. J. Parasitol.* 42, 255-237). These data provided information on potential gene targets for silencing. After optimising protocols to deliver dsRNA to *P. thornei* by ‘soaking’, the results showed that *P. thornei* and *P. zae* are both highly amenable to gene silencing. Following soaking of RLNs in solutions containing dsRNA to two genes involved in nematode movement, and culture for five week on ‘mini’ carrot discs, there was a 77-81% reduction in nematode replication (Tan, J-A.C.W. et al 2013, *Experimental Parasitology* 133, 166–178). These results were confirmed and extended by challenging transgenic wheat and sugarcane plants expressing dsRNA to target genes with root lesion nematodes.

27 Comparative analysis of complete mitochondrial genome sequences confirms independent origins of plant-parasitic nematodes

J.-K. Park¹, T. Sultana², J. Kim¹, S.-H. Lee¹ and T. Kim¹

¹*Program in Cell Biology and Genetics and Department of Parasitology, College of Medicine, Chungbuk National University, Cheongju 361-763, South Korea*

²*Department of Biological Sciences, Inha University, Incheon 402-751, South Korea*

Corresponding author: jkpyou@chungbuk.ac.kr

The nematode infraorder Tylenchomorpha (Class Chromadorea) includes plant parasites that are of agricultural and economic importance, as well as insect-associates and fungal feeding species. Among tylenchomorph nematodes, the superfamilies Tylenchoidea and Aphelenchoidea represent the largest assemblages of plant-parasitic chromadorean members. Monophyletic grouping and/or phylogenetic positions of these two superfamilies within chromadorean nematodes have been the topic of debate over many decades. We investigated phylogenetic relationships of the Tylenchoidea and Aphelenchoidea among other members of chromadorean nematodes based on comparative analysis of complete mitochondrial genome data, including three newly sequenced complete genomes from *Bursaphelenchus mucronatus*, *B. xylophilus* (Aphelenchoidea) and *Pratylenchus vulnus* (Tylenchoidea). Phylogenetic hypotheses for these mitochondrial genomes, based on different tree-building methods, did not support their monophyly: Aphelenchoidea was positioned basal to the Rhabditomorpha+Diplogasteromorpha+Ascaridomorpha+Panagrolaimomorpha clade, and Tylenchoidea was found to be the most basal taxon of the Chromadorean clade. Comparison of gene arrangement data corroborated the phyletic separation of these two groups: Similar gene arrangement patterns are found among the sampled species of the Rhabditomorpha /Diplogasteromorpha/ Ascaridomorpha/ Panagrolaimomorpha clade and aphelench species, with some minor exceptions. In contrast, only a single block (*rrnL-nad3*) is shared between aphelench (*Bursaphelenchus* spp.) and all three tylench (*P. vulnus*, *Radopholus similis* and *Heterodera glycines*) species. Additional mitochondrial genome sequences from as yet unsampled taxa will provide useful information for better characterizing the deep node phylogeny and mitochondrial genome evolution of nematodes.

28 Comparison of transcriptome pre- and post-parasitic stages of the nematode *Heterodera avenae*

D. Yang, C.L. Chen, S.S. Liu, Q. Yang and H. Jian

Department of Plant Pathology, China Agricultural University, Beijing, 100193, China

Corresponding author: hengjian@cau.edu.cn

As a worldwide plant pathogen, *Heterodera avenae*, is an obligate parasite in cereal crops. In China, the occurrence of *H. avenae* had distributed in 13 provinces and about 4 million hectares wheat fields were infested, and reduced the grain yield in significant proportions. However, a lack of genomic information and less genes information in the public databases has hindered the comprehensive elucidation of the molecular mechanisms coordinating its parasitism and pathogenicity. Using 454 Flx+ pyrosequencing, we analyzed the transcriptome of *H. avenae* and collected totally 1,066,719 reads including 551,935 from the pre-parasitic stages and 514,784 from the post-parasitic stages. These were assembled into 10,841 contigs with a mean length of 1,440 bp, among which 2892 contigs were differentially expressed between pre- and post-parasitic stages, and remained 71,401 singletons with an average length of 430 bp. Homology searches revealed that 59% of all contigs had significant matches with annotations to NCBI Nr database. In addition, 2855 (26%) and 1856 (17%) of those contigs were functionally classified using GO hierarchy and KEGG pathway respectively. The post-parasitic up-regulated genes mainly enriched in the some metabolism pathways such as Amino Acid Metabolism, Carbohydrate Metabolism, Lipid Metabolism and Biosynthesis of Other Secondary Metabolites, because life cycle of parasitic nematode turned into a more active phase after infection. We also identified some effectors in pre-parasitic stages expressed highly such as mimicking plant annexin 4F01, plant cell wall degradation enzymes GHF5 beta-1,4-endoglucanase, pectate lyase and so on. Furthermore, a lists of new putative effectors were found and under investigation.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

29 Identification of an annexin-like parasitism gene from cereal cyst nematode, *Heterodera avenae*

C.L. Chen, S.S. Liu, D. Yang, P. Liu, Q.X. Guo, Q. Yang and H. Jian

Department of Plant Pathology, China Agricultural University, Beijing, 100193, China

Corresponding author: hengjian@cau.edu.cn

Parasitism genes encoding secreted effector proteins of plant parasitic nematodes play important roles in facilitating parasitism. An annexin-like gene was isolated from the cereal cyst nematode *Heterodera avenae* with the most similarity to *annexin 2* which encodes a secreted protein of *Globodera pallida*. Southern blotting revealed that there are at least two copies of *annexin* in *H. avenae*. This identified *Ha-annexin* encodes a predicted 326 amino acid protein containing four conserved annexin domains. The protein has no N-terminal secretion signal peptide predicted by SIGNAL P 4.0, which is the same with ANNEXIN 2 of *Globodera pallida*. However, *in situ* hybridization showed that *Ha-annexin* transcripts exclusively expressed in the subventral gland cells of the pre-parasitic second-stage juveniles, which indicated that Ha-ANNEXIN is probably a secreted effector protein as ANNEXIN 2 of *Globodera pallida*. Quantitative real-time RT-PCR analysis confirmed that *Ha-annexin* was up-regulated in the parasitic second-stage juveniles, correlating with the time when feeding cell formation is initiated. When transiently expressed in onion epidermal cells, Ha-ANNEXIN was localized in the whole cell. Together, these results suggest that *Ha-annexin* most likely encodes a secreted effector protein that contributes to the early parasitic-stage process of *H. avenae*. The detail functions of this gene need to be analyzed in future.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

Session VII: Biological control and IPM

Chairmen: Prof.Joong-Ki PARK , Prof. Jingwu ZHENG

30 Control Options against the Cereal Cyst Nematodes *Heterodera filipjevi* in Turkey

Amer Dababat, G. Erginbas-Orakci, Ali Osman Sari, H.J Braun, and A. Morgounov

Amer Dababat: CIMMYT Global Wheat Program, Ankara-Turkey, a.dababat@cgiar.org

Gul Erginbas Orakci: CIMMYT Global Wheat Program, Ankara-Turkey, G.Erginbas@cgiar.org

Ali Osman Sari: Directorate of Research and Policy, Field Crops Research Department Ankara, Turkey, aosari@tagem.gov.tr

Hans Braun: CIMMYT Global Wheat Program, Mexico, h.j.braun@cgiar.org

Alexey Morgounov: CIMMYT Global Wheat Program, Ankara-Turkey, a.morgounov@cgiar.org

Corresponding author: a.dababat@cgiar.org

Soil borne pathogens including the Cereal Cyst Nematode (CCN) caused by *Heterodera* species are a threat facing cereal production systems worldwide and considered one of the major limiting cereal productions in rainfed regions among the plant parasitic nematodes. Damage exerted root system; therefore nematode damage is especially severe under rainfed growing conditions in more arid areas where water stress is regularly present. The most preferable method to control the CCN is through the use of genetic resistance. Resistance is biologically effective, economically acceptable, and environmentally friendly. However, up to now, resistance has only been identified against one of the CCN nematodes, *H. filipjevi* in Turkey and foreign wheat germplasm though this resistance is not yet present in high yielding cultivars. Resistance to the other nematodes in the CCNs complex is still being sought. Therefore, alternative approaches limiting the damage caused by CCN to wheat are needed. The soil borne pathogens program at CIMMYT Turkey established different control options under the umbrella of the Integrated Pest Management strategy to control cereal nematodes including the use of endophytic microorganisms and the use of seed treatment through establishing bilateral project with leading institutes. As a result of screening wheat germplasm against the CCN hundreds of moderately resistant to *H. filipjevi* in winter wheat germplasm are available now and many of them were distributed to international collaborators and they are being implemented in the breeding programs. The preliminary results of using seed treatments showed that seed treatment of wheat susceptible germplasm gave up to 47% reduction in number of *H. filipjevi* cyst per plant but did not reduce the number of cyst in the resistant germplasm since the cyst number was low and no room to decrease it further.

Keywords: Endophyte, *Heterodera*, Resistance, Seed treatment

31 Occurrence and Development of the Cereal Cyst Nematode (*Heterodera avenae*) in Shandong, China

Shuang Yu Xu, College of Plant Protection, Shandong Agricultural University, Taian 271018, China; and Hai Yan Wu, Agricultural College of Guangxi University, Nanning 530004, China; Jing Liu and Jian Luo, College of Plant Protection, Shandong Agricultural University, Taian 271018, China; and De Liang Peng, State Key Laboratory for Biology of Plant Diseases and Insect Pests, Chinese Academy of Agricultural Sciences, 100193 Beijing, China

Presenting author: E-mail address: whyzxb@yahoo.com.cn (H.Y.Wu)

Corresponding author: dlpeng@ippcaas.cn

The cereal cyst nematode (CCN) has been found in 16 provinces of China, including the Shandong winter wheat region. This study investigated dynamic changes in *Heterodera avenae* in root and soil for two consecutive years in a field experiment. Wheat roots were sampled during growing season, and *H. avenae* in wheat root and soil were counted. The results determined that the nematode densities in wheat root and soil of the two tested varieties had the same patterns over a two year period. Juvenile numbers in wheat roots were greatest during April when soil temperature was between 13–20.5°C. Cysts in rhizosphere soil increased significantly when new cysts were formed after Zadok47 (Booting Stage) ($P < 0.05$). There was a second-stage juveniles (J2) peak during Zadok 28 and 30 period, the number of J2 at the Zadok 13 (seedling stage) was the lowest. Our results provide important information indicating that the J2 population in root and soil increased after the wheat winter dormancy, which may provided valuable insights into an approach for integrated management of cereal cyst nematode, e.g. applying chemicals to kill J2 with irrigation after wheat winter dormancy.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

32 The researches of *Heterodera glycines* and resistant soybean germplasm in China

Yuxi Duan, Lijie Chen, Xiaofeng Zhu, Yuanyuan Wang, Xiaoyu Liu, Fang Wang, Dong Wang, Shuan Li
Nematology Institute of Northern China, Shenyang Agricultural University, Shenyang 110866, China

The soybean cyst nematode (*Heterodera glycines*, SCN) is a destructive pest of *Glycine max* (soybean). Five SCN samples from different areas were collected from China, the population P1 is race 1, HG-type is 2,5,7; the population P2 is race 2, HG-type is 1,2,5,7; the population P3 is race 3, HG-type is 5,7; the population P4 is race 4, HG-type is 1,2,3,5,7; the population P5 is race 5, HG-type is 1,3,4,7. There are more than 15 million hectares of soybean infested by SCN distributed in the provinces of northern of Yangtze River. And the most serious area of SCN is the west of Northeast of China. More than 100 resistant cultivars to SCN race 3 were identified from 15000 varieties. The resistance mechanism underlying *Heterodera glycines* infection is complicated.

Grouped and deducted the resistant genes from the 67 resistant soybean cultivars (Liaodou10 and Lee68 as the susceptible control) by the interorganismal genetics concept and greenhouse identification technique. According to known resistant genes, unknown resistant genes were deducted for the other cultivars. The results suggested that the cultivars tested were classified 7 groups. Resistant genes to at least two races of *H.glycines* were included in each cultivar. Huipizhi black soybean, Wuzhai black soybean, PI437654, Pingdingshan small black soybean and Black soybean (8498) included more than one resistant gene. Those cultivars were better resistant plasmas in the all tested varieties. The results provided to the breeders more information for selecting resistant soybeans and would accelerate breeding procedure for more new resistant varieties to *H.glycines*.

The suppressive subtractive hybridization (SSH) libraries were constructed to identify the differential gene expression profiles. RNAs were extracted from roots tips of resistant-nematode *G.max* Xiaoliheidou at 12h, 24h, 36h, 48h, 72h post inoculation and non-inoculation, respectively. A total of 166 expressed sequence tags (ESTs) spliced by CAP3 software were obtained after differential screening and northern blot. Those annotated genes by BLAST from the forward and reverse libraries were classified into nine functional categories including plant-pathogen interaction, stress and defense related, transcription regulation, signal transduction, transport, metabolism, cell cycle and DNA processing, cell component, protein synthesis and nucleotide binding. There were 16 differentially expressed genes were confirmed by quantitative real-time PCR. The trends of genes expression profiles were elevated in SCN-infected roots compared to uninfected roots including glutathione S-transferase, syringolide-induced protein, glucose-6-phosphate-dehydrogenase, nodulin-26, S-adenosylethionine synthetase, isoliquiritigenin 2'-O-methyltransferase-like, cinnamoyl coA reductase-like protein, Ring-H2 finger protein, glutathione peroxidase. The expression of genes like cyclin-dependant kinases and serine hydroxymethyltransferase1 were induced to decrease after infection. Genes encoding lipoxygenase, phenylacetaldehyde reductase, MYB transcription factor, histone showed to be up-regulated or down-regulated at different time point.

Acknowledgement: This research was supported by Special Fund for Agro-scientific Research in the Public Interest (200903040-03).

33 Biocontrol of cereal cyst nematode by two bacteria agents

Jie Zhang, H.-X. Yuan, Y.-H. Li, B.-J. Sun, X.-P. Xing and H.-L. Li

Department of Plant Pathology, Henan Agricultural university, NO.95 Wenhua Road, Jinshui District, Zhengzhou, 450002, P. R. China

Corresponding author: honglianli@sina.com

The cereal cyst nematodes (CCN) are recognized as one of the most important disease on cereals, which has caused significant damage and yield loss in Huanghuai wheat production area of China. With more attention on human health and environment, the problems caused by chemical control have led to increasing interest in the use of biological microorganisms.

In order to seek for biocontrol agents to this disease, 33 strains of bacteria were isolated from the cysts in the diseased fields of Henan province. Their biocontrol abilities to CCN were tested in pots and it indicated that the two isolates (strain 09B18 and strain 09X01) showed good effect to this disease. This study was done to investigate the role of the bacteria 09B18 and 09X01 as biocontrol agent against CCN. They were identified by morphological classification, physiological biochemical characteristic analyses and molecular identification. The bacterial colony features was observed and gram staining was conducted by both the human eye and scanning electron microscope to make preliminary appraisal. The physiological characteristics were studied according to *Bergey's Manual of Systematic Bacteriology* such as carbon utilization, voges-proskauer test, acid production of xylose. As for molecular identification, DNA was extracted and purified by "Dneasy Tissue Kit" and 16S rRNA gene was amplified by PCR and the primers were 27F(C) AGAGTTTGATCCTGGCTCAG and 1492R(C) TACGGCTACCTTGTTACGACTT. Also, the sequences were submitted on genebank and registration number were obtained. By dressing the seed with the bacteria suspension of 10^8 cfu/ml on beef extract peptone medium, the nematocidal potential of the two strain was tested in the greenhouse in 2011, and it was tested in the diseased fields of Xuchang city in Henan in 2012. Their activity was compared with that of 60% Avermectin (6ml/Kg). At the same time, the effect of the treatment on the growth parameters of wheat plant was also examined. The biocontrol mechanism was studied through the mortality of second stage juveniles (J2) under different exposure times (24, 48 and 72 hours) and the inhibition of eggs hatching in the cell suspension of 10^8 cfu/ml and filtrates respectively, with sterile water as control.

Based on a comparative 16S rRNA gene sequence analysis, strain 09B18 (sequence number is FJ982658) is closely related to *Bacillus cereus* (99%) and 09X01 (sequence number is HM854372) is close to *Achromobacter xylosoxidans* (99%) respectively. Combined with morphological and physiological characteristics, 09B18 strain was identified as *Bacillus cereus*, which was first report as biocontrol agents of cereal cyst nematode, and strain 09X01 was determined as *Achromobacter xylosoxidans*, which was first report as biocontrol agent of cyst nematode. In 2011, the cysts decreasing rates of the treatment with 09B18 and 09X01 agent were 75.87% and 70.21% in the greenhouse. It was 37.54% and 35.55% respectively in Xuchang diseased fields in 2012. While the cysts decreasing rates of 60% Avermectin (6ml/Kg) in pot and the field was 64.14% and 54.18%, respectively. At the same time, the bacteria 09B18 can increase the fresh weight and the output of wheat plant obviously. The adjusted mortality of 09B18 filtrate on second stage juveniles at 72h reached 100% and 09X01 filtrate reached 99.5%. The filtrates of the two bacteria showed high insecticidal activity to juveniles, while the suspension of the bacteria cells was rarely to kill second stage juveniles. The percentage of egg hatching with 09B18 filtrate was 20% after 30 days while the rate in sterilized water was 83%. Thereafter, the two bacteria can be supposed as potential biocontrol agents of cereal cyst nematode of wheat by drawing them through integrated pest management programs.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

34 First report of cystoid nematodes in China with notes on *Cryphodera sinensis* n. sp.

(Nematoda: Heteroderidae)

Kan Zhuo¹, HongHong Wang¹, Weiming Ye², Deliang Peng³ and Jinling Liao^{1*}

¹Laboratory of Plant Nematology, South China Agricultural University, Guangzhou 510642, China;

²Nematode Assay Section, Agronomic Division, North Carolina Department of Agriculture & Consumer Services, 4300 Reedy Creek Road, Raleigh, NC 27607, USA;

³State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China

Corresponding author: jliao@scau.edu.cn

The family Heteroderidae of plant-parasitic nematodes is a group of important plant parasites containing cyst nematodes (or cyst-forming nematodes) and cystoid nematodes (or non-cystforming nematodes), but less work on Heteroderidae has been done in Chinese tropical and subtropical areas. During the past three years, surveys for the presence of Heteroderidae nematodes in Chinese tropical and subtropical regions have been done. Besides some *Heterodera* species belonging to cystnematodes have been identified, three *Cryphodera* populations belonging to cystoid nematodes were found by using comparative morphological, morphometric and molecular studies. Among these three populations, one from Hunan province has unique morphological characters, SSU, LSU D2D3 and ITS rRNA sequences, and it was identified as a new species, namely *Cryphodera sinensis* n. sp. In addition, the phylogenetic trees based on LSU D2D3 and rDNA-ITS showed that the other two *Cryphodera* populations from Guangxi and Hainan province are in a strong supported monophyletic clade with other *Cryphodera* species. Because only second-stage juveniles were obtained from these two *Cryphodera* populations, it is impossible to identify what kind of species the two *Cryphodera* populations are. In China, cystoid nematodes have never been reported to date. This is the first record of the cystoid nematode *Cryphodera* in China.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

35 Diagnosis of *Cryphodera brinkmani* Intercepted on *Juniperus chinensis* Imported into China from Thailand

Hai LONG, Yi-Nong LI, Fang-Rong LI

Shenzhen Entry-exit Inspection and Quarantine Bureau, Shenzhen 518045, China

The non-cyst forming heteroderid nematode *Cryphodera brinkmani* was detected in roots of Chinese juniper (*Juniperus chinensis*) imported from Thailand. Lots of larvae and males were found, fewer females. Morphology and morphometrical traits of the intercepted population on this new host for *C. brinkmani* were in agreement with the original description, except for some minor differences on male morphology. Molecular data for this species were obtained using D2-D3 expansion regions of 28S rDNA. The phylogenetic relationships of this species with other representatives of non-cyst and cyst-forming Heteroderidae using 28S rDNA were presented and indicated that *C. brinkmani* clustered together with other *Cryphodera* sp. Analysis of morphology and molecular biology confirmed and supported the species identifications. According to our knowledge, it is the first time this nematode was intercepted on new host from new country.

Key word: intercept; *Cryphodera brinkmani*; *Juniperus chinensis*; Thailand

36 Temperature-manipulated development of novel cellular and mycelium stages of

Pasteuriapenetrans

X.X. Bu, H. Jian, Z.X. Chen, X.Z. Liu and Q. Liu

Department of Plant Pathology, China Agricultural University, Beijing, 100193, China

Corresponding author: hengjian@cau.edu.cn

The development of *Pasteuria penetrans* in root-knot nematodes had been further studied using light and scanning electron microscopy. Some rod-like bacilli were observed when root-knot nematodes infected with *P. penetrans* were cultured for 12 days at 25°C/35°C 10 h dark/14 h light. Rod-like bacilli were 0.6-0.9 µm in length and about 0.6-0.9 µm in diameter and some of them accumulated to form cellular masses. Numerous of thalli had produced two days later and gathered around the metacopu or the intestine of nematode. At this time, mycelia like zingiberhitherto not documented were observed under scanning electron microscopy. The mycelia like zingiber should be long to vegetative thalli and they were still detected after 700 accumulated degree days. It suggested that vegetative growth and differentiation may simultaneously occur in most thalli. The development of thalli would be stopped if they were cultured at $16 \pm 1^\circ\text{C}$. This status can remain no less than 30 days. *P. penetrans* can recover to develop and produce mature endospores if they were transferred to normal condition. The development rate after transferred to the natural condition was still the same as the control culture at the same condition all along. Comparing the yield of spores developing from low temperature treated mycelia with the control, and the result indicated there was no significant different. But the sporulation of sporangia would not be stopped, even if they were cultured at $16 \pm 1^\circ\text{C}$.

37 QTL analysis of gene *RKN2* in *Gossypium barbadense* which clusters with gene *rkn1* in *G. hirsutum* for transgressive nematode resistance

Congli¹ Wang*, M. Ulloa², and P.A. Roberts¹

¹University of California, Riverside, CA 92521

²USDA-ARS, Lubbock, TX 79415

Current address: Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Harbin, Heilongjiang Province, 150081

A transgressive factor *RKN2* from susceptible *Gossypium barbadense* Pima S-7 to the root-knot nematode (RKN, *Meloidogyne incognita*) interacts with the resistance gene *rkn1* in *G. hirsutum* AcalaNemX to produce higher resistance. The two genes are clustered together linked to SSR marker CIR316, and QTL analysis in an F_{2:7} (Pima S-7 x AcalaNemX) population confirmed marker CIR316 on the telomeric region of chr11 contributed 51% of resistance to both root-galling and nematode egg production. QTL analysis of the test cross AcalaNemX x F₁ (Pima S-7 x Acala SJ-2) indicated the allele of marker CIR316 from Pima S-7 on chr11 contributed 29% and 27% of resistance to root-galling and nematode egg production, respectively, in the presence of *rkn1*. When susceptible Pima S-7 was crossed with susceptible Acala SJ-2, transgressive resistant lines were found in F₂ progenies. QTL analysis of F₂ (Pima S-7 x Acala SJ-2) suggested that the allele of marker CIR316 on chr11 contributed 15% and 22% of resistance to root-galling and nematode reproduction, respectively. There was no contribution on homoeologous chr21 with different alleles from those on chr11. BAC sequences from marker CIR316 close to RKN resistance genes contained two copies of resistance gene analogs (RGA), one of the RGA sequences (3148 bp) of CIR316 on chr11 had 83% identity with another RGA (3375 bp) of CIR316 on chr21. These sequence comparisons provided further insight into the organization and molecular evolution of the RKN-resistance gene cluster on chr11 and its homoeolog chr21. Based on these and previous findings, transgressive segregation is common in cotton and efforts to identify novel phenotypes for biotic and abiotic stress resistance traits among progenies developed from stress susceptible or sensitive parent combinations are worthy of increased attention in plant breeding programs.

General Abstracts

38 Pathotype Stability of *Heterodera avenae* from Shanxi, China and suggestion on revision of Pathotype characterization system

Jingwu Zheng¹, Hongli Shi¹, Deliang Peng²

¹Institute of Biotechnology, College of Agric. & Biotechnology, Zhejiang University, Hangzhou 310058, China

²Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193

Corresponding author: jwzheng@zju.edu.cn

Cereal cyst nematode, *Heterodera avenae*, is one of the most important limit factors for wheat production in China. The present pathotype characterization system for *H. avenae* was put forward by S. Andersen and K. Andersen in 1981, which based on the reaction of nematode population to 23 cultivars or lines of cereals (International Test Collection, ITC), including 11 of Barley, 7 of Oats, and 5 of wheat. All of which were divided into two group A and group B. In 1997, one cereal cyst nematode population from Taigu, midland of Shanxi, China were characterized based on the reaction to group A. It was found that all the barley and oats cultivars were resistant, only Capa, Festigyay, and Aus 13807 of wheat were susceptible. During 2011-2012, two *H. avenae* populations from Wenxi and Yuncheng, south of Shanxi were characterized for their pathotype based on the reaction to the ITC. All the data are similar to that made in 1997 to Tagu population. It showed that the pathotype of *H. avenae* in Shanxi is relatively stable. Because of the wide distribution in China, and most of ITC are resistant to the Chinese *H. avenae* populations. We do think it is good to add some Chinese resistant cultivars and omit some of foreign resistant cultivars or lines in ITC. That will be useful for monitor the pathotype variation of *H. avenae* in China.

Key words: *Heterodera avenae*, Pathotype, International Test Collection.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

39 Research on infection and development dynamics of *Heterodera avenae* to spring wheat in Qinghai province

Sheng-ying HOU, Gui ZHANG, Xing WANG, Ai-ling WANG

Institute of plant protection, Qinghai Academy of Agricultural and Forestry Sciences, Qinghai University, Xining, 810016

The infection period, development progress, dynamics and generation of *Heterodera avenae* on spring wheat in Qinghai province was carried out by using seedling observation combined by field investigation. The results show that 2 instar larvae hatched out of the overwintering cyst in soil infected the wheat seedling roots by the rise of temperature. Since the middle of April, the larvae completed three stages from two age to four age in wheat root late April to early July and the white female adults exposed on the root surface In July. From August to September the new cyst fallen into the soil, and it's for Wintering period from the middle of October to next April. *Heterodera avenae* occurs one generation during the whole growth period of spring wheat.

Key words: *Heterodera avenae* Woll.; infection; development; dynamics

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

40 The dynamics of *Heterodera avenae* in winter wheat in Hebei province

X.-H. Li, J. Ma, B. Gao, R.-Y. Wang and S.-L. Chen

Institute of Plant Protection, Hebei Academy of Agricultural and Forestry Sciences/ IPM Center of Hebei Province/ Key Laboratory of Integrated Pest Management on Crops in Northern Region of North China, Ministry of Agriculture, P. R. China, Baoding 071000, Hebei Province, China
Corresponding author: chen_shulong@tom.com

Cereal cyst nematode (*Heterodera avenae*) has become the main disease in wheat production in China and even worldwide. Understanding the dynamic of cereal cyst nematode in the field is important for the control of the disease. Three surveys were conducted in the main winter wheat production area in Hebei from October, 2009 to September, 2012. The results showed that few J2 were detected in soil in wheat sowing period (October 6th), and it reached the peak (12.3-18.6 J2/100ml soil) in late November (23th to 27th, Nov.) before the soil freeze, then the number of J2 dropped to 0.1 J2/100ml soil in 23rd of December and no J2 isolated in soil layers (0-14 cm) in January. Afterwards the number of J2 in soil increased again with the soil temperature increasing, and reached the peak in early April with 52- 65 J2/100ml soil, it decreased to 1.3 J2/100ml soil in May 12th. No J2 isolated from early June. Very low number of J2 with 0.4-3 J2/100ml soil could be extracted in the soil from July to September. J2 could penetrate root from October 15 (0.7 J2/plant) to November 10th (0.3 J2/plant), no obvious peck found before soil freeze and the nematodes stop to penetrate during soil freeze. J2 started to penetrate root again from end of February and the higher number of J2 with 59-102 J2/plant were found from 6th-14th of April, then the number of J2 penetrated to the root decreased. Accordingly, third stage juveniles formed between October and November, and peaked between late April and early May. The white females were detected from mid of May and higher number of cyst formed in late May. The eggs in the cyst from June to end of July were mainly in the embryonic development stage. In summary *H. avenae* occurs one generation per year in wheat in Hebei province and the main damage period is from late March to early April in spring.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

41 Pathotype characterization of the cereal cyst nematode *Heterodera avenae* in Beijing, China

Jiangkuan Cui, WenKun Huang, Wenting He, Ling'an Kong, DeLiang Peng

State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China.
Corresponding author: dlpeng@ippcaas.cn

The cereal cyst nematode (*Heterodera avenae*) has caused huge yield loss on wheat crops. Pathotype diversity study could offer support for wheat planting, diseases control, breeding for disease resistance and choices of varieties. The pathotype of *H. avenae* in Beijing, China was identified using 23 standard international differentials and 4 local cultivars Wenmai19, Aikang58, Taikong6, Zhongyu in 2011 and 2012, respectively. Three differential barleys La Estanzuela, Dalmatische and Varde were susceptible. The six oats were resistant to the tested nematode population. The wheats Iskamisch K-2-light and Psathias were resistant while the Capa, Loros, AUS 10894 and 4 local cultivars were all susceptible. The results showed the population avirulent to Ortolan (Ha 1), Hetar, Sun II. According to the character item, which showed the Caiyu CCN population virulence pathotype classified to be group Ha 31.

Acknowledgments: This study was supported by the Special Fund for Agro-scientific Research in the Public Interest (No. 200903040)

42 Chemotaxis of *Ditylenchus destructor* to extracts from sweet potato

J. Ma, X.-H. Li, R.-Y. Wang, Y.-L. Geng and S.-L. Chen

Institute of Plant Protection, Hebei Academy of Agricultural and Forestry Science / IPM centre of Hebei Province/ Key Laboratory of IPM on Crops in Northern Region of North China, Ministry of Agriculture, 437 Dongguan street, Baoding, Hebei 071000, China

Corresponding author: chenshulong@gmail.com

Ditylenchus destructor Thorne could cause severe damage to sweet potato, potato and other host plants in China. Host plant phytochemicals play important roles in the nematode behavior, thus identification of the compounds that attract the nematodes to the plant is important for the nematode control. In the present study, experiments were carried out to investigate chemotaxis of *D. destructor* to different host plant and determine the attraction intensity of different host plants. Sweet potato was consistently and significantly much more attractive than potato and carrot in all these assays. Compared with the stems, leaves and storage roots of sweet potato, stems showed the significantly strongest attraction ability to nematodes and our result showed that the attractant from sweet potato was stable to heat. Differences in the relative attraction to *D. destructor* among sweet potato cultivars were also studied, the results showed that the stems of susceptible cultivars Lizixiang, Jishu 98, Shangshu 19 were significantly more attractive than the resistant cultivars Xushu 25, Xushu18, Jishu 17-52; in contrast, there were no significant difference of the storage roots among different sweet potato cultivars. Furthermore, the compounds of sweet potato were extracted and studied in relation to the host-finding behavior of the nematode. The fractions extracted with butyl alcohol from stems were significantly more attractive than extractions by water, alcohol, petroleum ether and ethyl acetate. The results of this study suggest that extracts from sweet potato can potentially be used as baits in a trap for the control of *D. destructor* in the field.

Key words: *Ditylenchus destructor*, compounds, attraction, chemotaxis

43 The detection of cyst-forming nematodes on Poaceae in China

D. Wang, X.-F. Zhu, F. Zhu, Y.-Y. Wang, D. Liu, L.-J. Chen and Y.-X. Duan*

Nematology Institute of Northern China, College Plant Protection, Shenyang Agricultural University, Shenyang, 110866, P. R. China.

Corresponding author: duanyx6407@163.com

Members of cyst nematodes are among the most important plant-parasitic nematodes in agriculture, which can infect plant hosts from Poaceae. During a survey from 2010 to 2012, some populations of cyst nematodes parasitized Poaceae were detected by the sieving-decanting method on the roots and in rhizosphere soil in China. Based on both morphological and molecular identification, four species including *Heterodera avenae*, *H. filipjevi*, *H. elachista* and *Cactoderaestonica* were indentified. *H. filipjevi* from Henan province and *H. avenae* from Anhui, Hebei, Henan, Shandong and Shanxi provinces infected wheat, which belong to ‘Avenae’ group. *H. elachista* on rice from Hunan province belongs to the ‘Cyper’ group, while two populations of *Cactoderaestonica* from Liaoning province were recovered on *Setaria viridis* (L.) Beauv, one of grass from Poaceae. This work was supported by the Special Fund for Agro-scientific Research in the Public Interest 200903040-03.

Acknowledgement: The research was supported by Special Fund for Agro-scientific Research in the Public Interest in China (No.200903040)

44 Biological characteristics of *Heterodera elachista* on rice in Hunan, China

DING Zhong¹, Wang Shuinan¹, NAMPHUENG Janthathang¹, Peng Deliang², Huang Wenkun²

¹ College of Plant Protection, Hunan Provincial Key Laboratory for Biology and Control of Plant Disease and Insect Pests, Hunan Agricultural University, Changsha 410128, China.

² The Key Laboratory for Biology of Insect Pests and Plant Disease, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China.

Corresponding author: dingzx88@aliyun.com

The rice cyst nematode (*Heterodera elachista*) is an important pest in hilly rice fields of Hunan Province, China, and has been previously identified from rice fields in Japan and Iran. *H. elachista* can decrease yield by 7–19%, and has the most severe impact during the later stages of plant growth. Biological characteristics of *Heterodera elachista* were investigated by using artificial inoculations with second-stage juveniles in laboratory and periodical sampling in rice field. The optimum temperature range for hatching of J2 from cysts of *H. elachista* was 28 to 32 °C, and root diffusate of rice, leachates of rice soil and twenty-time rice root juice stimulated emergence of J2 from cyst. The shortest life cycle at 30 °C from the infective juvenile to the emergence of second-stage juvenile is 18 days while from egg to egg stage is 22 days. The invasion and development of *H. elachista* were favored by relatively high temperatures, e.g. 28~35 °C. Root infection and yield loss caused by *H. elachista* on rice can be greatly reduced with appropriate management of irrigation water. *H. elachista* is unable to penetrate rice roots under anaerobic soil conditions.

Acknowledgement: This research was supported by Chinese Special R & D Fund for Public Benefit Agriculture (200903040).

45 Effect of Abamectin on cereal cyst nematode of wheat

Tao Wang, W. Zhao, W.-K. Huang, D.-L. Peng and R.-D. Qi *

Institute of Plant Protection and Agro-products Safety, Anhui Academy of Agricultural Sciences, No. 40 Nongkenan Road, Hefei, 230031 Anhui, China.

Corresponding author: rende7@126.com

Cereal cyst nematode (CCN) is globally and economically important in wheat production systems, and distributed widely in main wheat growing areas in China. In order to investigate the effect of Abamectin against CCN and its application method, the plot trials were conducted by soil treatment, seed processing and root irrigation with Abamectin emulsifiable concentrate (EC), controlled releasing Abamectin capsule suspension (CS) and Abamectin granules (GR), respectively. Three groups of results were obtained as follows: (i) the control efficacies of Abamectin against CCN by root irrigation were 22.3% and 31.4% with Abamectin EC (225 g a.i./ha) and Abamectin CS (225 g a.i./ha) before overwintering, respectively, and 73.6% and 70.4% during regreening stage of wheat, respectively. (ii) the control efficacies against CCN were 21.8%, 16.6% and 27.8% by soil treatment with Abamectin G, Abamectin EC and Abamectin CS before sowing, respectively. (iii) the efficacies against CCN were 23.4% and 52.4% by seed dressing with Abamectin EC and Abamectin CS at the ratio of 2% of seed weight, respectively. The results suggested that Abamectin was an available nematicide to control CCN, and taken together, seed processing with the controlled releasing Abamectin CS must be the most effective method considering the ease of operation and efficacy based on our study.

Acknowledgement: This study was sponsored by the special fund for Agro-scientific Research in the Public Interest (200903040).

46 Identification of two pharyngeal gland specific pectate lyases from the cereal cyst nematode *Heterodera avenae*

Haibo Long^{1,2}, Xiaochuan Gu¹ and Deliang Peng^{1*}

¹State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193;

²Key Laboratory of Pests Comprehensive Governance for Tropical Crops, Ministry of Agriculture, Environmental and Plant Protection Institute, Chinese Academy of Tropical Agricultural Sciences, Haikou, 571101, China

Corresponding author: dlpeng@ippcaas.cn

Parasitism genes encoding secretory proteins expressed in the pharyngeal glands of plant-parasitic nematodes play crucial roles in nematode parasitism of plants. Two pectate lyases genes (*Ha-pel-1* and *Ha-pel-2*) expressed in the pharyngeal glands of the sedentary cyst nematode, *Heterodera avenae*, were cloned from a Pre-parasitic second stage juveniles cDNA library, and the corresponding genomic DNAs were subsequently cloned. *Ha-pel-1* and *Ha-pel-2* consist of 1717 nucleotides encoding 521 amino acids and 1000 nucleotides encoding 328 amino acids, respectively. DNA gel blotting confirmed that these two genes were of nematode origin and present as members of a small multigene family. The deduced protein sequences HA-PEL-1 and HA-PEL-2 share only 27% identity and 46% similarity. Phylogenetic analysis revealed that they cluster in two different clades. Both of the predicted proteins have a putative signal peptide for secretion. *In situ* hybridization showed that the transcripts of *Ha-pel-1* and *Ha-pel-2* accumulated specifically in the two subventral gland cells of *H. avenae*. Moreover, RT-PCR showed that both genes were expressed in the migratory preparasitic stage although the level of expression between the two genes was different. These results indicate that HA-PEL-1 and HA-PEL-2 may be important enzymes early in the migration process.

Acknowledgments: This research was supported by the National Natural Science Foundation of China (No. 31201493) and by the Special Fund for Agro-scientific Research in the Public Interest (No. 200903040).

47 The distribution and rDNA-ITS analysis of Cereal Cyst Nematode (*Heterodera avenae*) in Shandong Province, China

Chong-Jun LIU^{1,2} Wen-Kun HUANG¹ Jiang-Kuan CUI¹ De-Liang PENG^{1*} Hong-Mei LI^{2*}

¹State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China

²College of Plant Protection, Nanjing University, Nanjing 210095, China;

Corresponding author: dlpeng@ippcaas.cn

The cereal cyst nematode (*Heterodera avenae*) is the most important nematode on wheat and cereal crops in China. The occurrence and distribution of *Heterodera* spp. were investigated by random sampling method from 19 counties of Shandong province. The species was identified as *Heterodera avenae* with the morphological and rDNA-ITS analysis. The cereal cyst nematode (*Heterodera avenae*) was detected from 84.2 percent samples collected from Linyi, Laiwu, Zibo, Weifang, Dongying, Weihai, Yangtai. The highest cyst and egg number existed in Dongying city and Weifang city, the lowest cyst and egg number existed in Linyi city and Yantai city. This survey results will be beneficial for making suitable management strategy and control measures.

Acknowledgement: This Research was supported by Chinese Special R & D Fund for Public Benefit Agriculture (200903040) and Ministry of Science and Technology (2009DFB30230).

48 Molecular characterization and functional analysis of a new acid phosphatase gene (*Ha-acp1*) from *Heterodera avenae*

Yan-Ke, LIU; WenKun Huang; Gaofeng Wang, Haibo, LONG; Huan, PENG Wenting, HE; DeLiang PENG *

State Key Laboratory for Biology of Plant Diseases and Insect Pests, Institute of Plant Protection, Chinese Academy of Agricultural Sciences, Beijing 100193, China.

Corresponding author: dlpeng@ippcaas.cn

For sedentary endo-parasitic nematodes, parasitism genes encoding secretory protein expressed in the subventral glands cells always play an important role during the early parasitic process. A new acid phosphatase gene (*Ha-acp1*) expressed in the subventral glands of the cereal cyst nematode (*Heterodera avenae*) was cloned and the characteristics of the gene were analyzed. Results showed that the gene had a putative signal peptide for secretion and *in situ* hybridization showed that the transcripts of *Ha-acp1* accumulated specifically in the subventral gland cells of *H. avenae*. Southern blot analysis suggested that *Ha-acp1* belonged to a multigene family. RT-PCR analysis indicated that this transcription was strong at the pre-parasitic juveniles. Knocking down *Ha-acp1* using RNA interference technology could reduce nematode infectivity by 50%, and suppressed the development of cyst. Results indicated that *Ha-acp1* could play an important role in destroying the defense system of host plants.

Keyword: *Heterodera avenae*, acid phosphatase gene, *Ha-acp1*, RNA interference

Acknowledgments: This research was support the National Natural Science Foundation of China (No. 31201493) and by the Special Fund for Agro-scientific Research in the Public Interest (No. 200903040).

Participant Lists of the 4th International Cereal Nematodes Initiative Workshop (August 22-24, Beijing china)

	Name	Title	Mailing Address	Email-address
1	Abdelfattah A.S. Dababat	Dr.	ICARDA-CIMMYT Wheat Improvement Program CIMMYT Global Wheat Program; CIMMYT; P.K.39 Emek 06511 Ankara TURKEY; Tel:90-312- 3448777; Fax: 90-312-3270798	a.dababat@cgiar.org
2	Ahmed A. M. I. Dawabab	Prof.	Plant Protection Department, College of Food and Agriculture Sciences, King Saud University, Egypt	adawabab@ksu.edu.sa
3	Andreas Westphal	Dr.	Institute for Plant Protection in Field Crops and Grassland, Messeweg 11/12; 38104 Braunschweig, Germany, Tel.: +49 (0)531 299 3929 Fax: +49 (0)531 299 3010	andreas.westphal@jki.bund.de
4	Alireza Ahmadi	Dr.	Agricultural Research and Natural Resources Centre of Khuzestan, Ahvaz, Iran	alirahmadi2000@gmail.com
5	Fateh Toumi	Mr.	Institute for Agricultural and Fisheries Research (ILVO), Burg. Van Gansberghelaan 96, 9820 Merelbeke, Belgium;	fateh.toumi@ilvo.vlaanderen. be
6	Fouad Mokrini	Mr	National Institute of Agricultural Research, (INRA), Km 9, 14000 Kenitra, Morocco. Institute for Agricultural and Fisheries Research, Plant, Crop Protection, Burg. Van Gansberghelaan 96, B-9820 Merelbeke, Belgium.	fmokrini@yahoo.com
7	Gul Erginbas Orakci	Ms	ICARDA-CIMMYT Wheat Improvement Program CIMMYT Global Wheat Program; CIMMYT (International Maize and Wheat Improvement Centre); P.K. 39 Emek 06511 Ankara TURKEY; Tel:90- 312-3448777; Fax: 90-312-3270798	G.Erginbas@cgiar.org
8	Halil Toktay	Dr.	Nigde University, Faculty of Agrucultural Science and Technology. Nigde Turkey	toktay@yahoo.com
9	Ibrahim Halil Elekcioglu	Prof.	Dean of Faculty of Agriculture, University of Cukurova 01330 Saricam, Adana, Turkey, Tel +90 533 360 80 47; Fax: +90 322 338 63 64	halile@cu.edu.tr
10	Joong-Ki Park	Prof.	Graduate Program in Cell Biology and Genetics/Department of Parasitology, College of Medicine, Chungbuk National University Cheongju 361-763, Chungbuk, Republic of Korea Tel:+82-(0)43-261-2843 (Lab.), CP:+82-(0)10-6431- 3735, http://molevol.chungbuk.ac.kr/	jkpyou@chungbuk.ac.kr
11	Maurice Moens	Prof.	Institute for Agricultural and Fisheries Research (ILVO), Burg. Van Gansberghelaan 96, 9820 Merelbeke, Belgium;	maurice.moens@ilvo.vlaander en.be
12	Michael Jones	Prof.	Prof. of Agricultural Biotechnology Director, WA State Agricultural Biotechnology Centre, Murdoch University, Perth WA Australia 6150 +61 8 9360 2424; +61 (0) 414239428	M.Jones@murdoch.edu.au

	Name	Title	Mailing Address	Email-address
13	Ricardo Holgado	Dr.	Norwegian Institute for Agricultural & Environmental Research, Plant Health & Plant Protection Division Department of Bacteriology, Virology & Nematology Høgskoleveien 7, N-1430 Ås, Norway Direct Phone + 47 916 827 03, www.bioforsk.no	ricardo.holgado@bioforsk.no
14	Richard W. Smiley	Prof.	Oregon State University Columbia Basin Agricultural Research Center P.O. Box 370, Pendleton, OR 97801, 48037 Tubbs Ranch Road, Adams, OR 97810 USA Tel: 541-278-4397 cell: 541-969-0910 fax: 541-278-4188	richard.smiley@oregonstate.edu
15	Saleh Alnadhary	Dr.	Plant Pathology - Nematode College of Food and Agriculture Sciences King Saud University, Tel. 00966568802471 Yemen	nadary44@gmail.com
16	Wenjing Pang	Dr.	Bayer Crop Science LP, BCS-SD-BioScience, ARD-ADIS-ADIS -ADIS, Morrisville, Tel: +1 919 461 6612,Fax: +1 919 461 6544; E-mail: wenjing.pang@bayer.com	wenjing.pang@bayer.com
17	Zahra Tanhamaafi	Dr.	Iranian Research Institute of Plant Protection, P.O. Box 1454, 19395 Tehran, Iran, Phone 98 21 22403012-16, Fax ++98 21 22403691	zahrata.maaafi@yahoo.com
18	Congli Wang	Dr.	University of California, Riverside, CA 92521, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, China; Tel:15045119624	clwangjb@yahoo.com
19	Baishuang	Prof.	Heilongjiang Academy of Agricultural Science,China; Tel:13304805716	bsyu100@163.com
21	Bingyan	Prof.	Institute of Vegetables and Flowers, Chinese Academy of Agricultural Sciences, China; Tel:13701013984	xieby@mail.caas.net.cn
22	Changlong Chen	Ph.D student	College of Agriculture and biotechnology,Chinese Agricultural University, China; Tel:15510028385	chenchanglong319@126.com
23	Dan Yang	Ph.D student	College of Agriculture and biotechnology,Chinese Agricultural University, China; Tel:62731171	yd13_0@163.com
24	Deliang Peng	Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences,China; Tel:13381056763	dlpeng@ippcaas.cn
25	Deyong Zhang	Prof.	Institute of Plant Protection, Hunan Province,China; Tel:13974848177	dyzhang78@gmail.com
26	Dong Wang	Ph.D student	Shenyang Agricultural University, China; Tel:15040031985	wangdong19852008@163.com
27	Doudou Huang	Student	College of Plant Science and Technology of Agricultural University, China; Tel:13720165436	huangdoudou23@163.com
28	Feixue Cheng	Asso Prof.	Institute of Plant Protection, Hunan Province,China; Tel:13974869232	cfx937207@126.com
29	Feng Liu	Asso Prof.	College of Plant Protection, Shandong Agriculture University, China; Tel:18653871817	fliu@sdau.edu.cn
30	Guokun Liu	Prof.	College of Plant Protection, Fujian Agriculture and Forestry University,China; Tel:13159432336	Liuguok@126.com
31	Hai Long	Asso Prof.	Shenzhen Entry-Exit Inspection and Quarantine Bureau, China	
32	Haibo Long	Dr.	Environment and Plant Protection Institute, Chinese Academy of Tropical Agricultural Sciences, China; Tel:13118916919	hb_long@yeah.net
33	Haiyan Wu	Prof.	Guangxi University,China; Tel:13077741203	wuha@sdau.edu.cn
34	Heng Jian	Prof.	College of Agriculture and biotechnology,Chinese Agricultural University, China; Tel:62731102	hengjian@cau.edu.cn

	Name	Title	Mailing Address	Email-address
35	Honghai Zhao	Prof.	Qingdao Agricultural University,China; Tel:13407601717	hhzhao@qau.edu.cn
36	Hongjie Li	Prof.	Institute of Crop Science, Chinese Academy of Agricultural Sciences, China; Tel:13552766063	lihongjie@caas.cn
37	Hongmei Li	Prof.	College of Plant Protection, Nanjing Agricultural University,China; Tel: 15951964113	lihm@njau.edu.cn
38	Hongxia Yuan	Asso Prof.	Agricultural University of Henan Province,China; Tel:13303830979	yhx2156@163.com
39	Hongyou Zhou	Prof.	Agricultural University of Inner Mongolia, China; Tel: 15847660292	hongyouzhou2002@aliyun.com
40	Hui Xie	Prof.	College of Natural Resources and Environment of Southchina Agricultural University, China; Tel:18675856173	xiehui@scau.edu.cn
41	Jianfeng Gu	Prof.	Ningbo Entry-Exit Inspection and Quarantine Bureau, China; Tel: 0574-87022839	gujif@nbciq.gov.cn
42	Jianhua Sun	Prof.	College of Chemistry and Lifesciences, Tianjing Normal University, China; Tel:13920899633	jianhuasun55@yahoo.com.cn
43	Jie Zhang	Ph.D student	Agricultural University of Henan Province, China; Tel: 15824860270	Zhangjie656@126.com
44	Jie Zhang	Ph.D student	College of Plant Protection, Shandong Agriculture University,China	
45	Jincheng Wang	Prof.	Tianjin Entry-Exit Inspection and Quarantine Bureau, China; Tel:13752253687	wangjincheng@yahoo.cn
46	Jingwu Zheng	Prof.	College of Agriculture and biotechnology,Zhejiang University, China; Tel: 0571-88982580	jwzheng@zju.edu.cn
47	Jinling Liao	Prof.	College of Natural Resources and Environment of Southchina Agricultural University, China; Tel:18928800937	jlliao@scau.edu.cn
48	Kan Zhuo	Assit Prof.	College of Natural Resources and Environment of Southchina Agricultural University, China; Tel:15913155915	zhuokan@scau.edu.cn
49	Lijie Chen	Prof.	Shenyang Agricultural University,China; Tel:13889390426	chenlijie0210@163.com
50	Ling-an Kong	Dr.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:62815611	lakong@ippcaas.cn
51	Rende Qi	Asso Prof.	Institute of Plant Protection, Academy of Agricultural Science,Anhui Province, China; Tel:13955172941	rende7@126.com
52	Ruyu Yao	Ph.D student	Yunnan Agricultural University, China; Tel:15288363430	yaoruyu@126.com
53	Shulong Chen	Prof.	Institute of Plant Protection, Hebei Academy of Agriculture and Forestry Sciences, China; Tel:13703220581	chenshulong@gmail.com
54	Songbai Zhang	Assit Prof.	Institute of Plant Protection, Hunan Province,China; Tel:13637427795	
55	Wenkun Huang	Asso Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences,China; Tel:13522412250	wkhuang2002@163.com
56	Wenting He	Assi Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:62815611	beautydaji@sina.com
57	Xiangxia Bu	Dr.	College of Agriculture and biotechnology,Chinese Agricultural University, China; Tel:15652168609	buxx83@163.com
58	Xianqi Hu	Prof.	Yunnan Agricultural University, China; Tel:13808701887	xqhoo@126.com

	Name	Title	Mailing Address	Email-address
59	Xiaofeng Zhu	Asso Prof.	Shenyang Agricultural University,China; Tel:13516039056	Syxf2000@163.com
60	Xinqiu Tang	Asso Prof.	Institute of Plant Protection, Hunan Province,China; Tel:13298691789	
61	Xiuhua Li	Assit Prof.	Institute of Plant Protection, Hebei Academy of Agriculture and Forestry Sciences, China; Tel:13472319597	lxh_727@yahoo.com.cn
62	Xuan Wang	Asso Prof.	College of Plant Protection, Nanjing Agricultural University, China; Tel:13813978667	xuanwang@njau.edu.cn
63	Yanhua Wen	Prof.	College of Natural Resources and Environment of Southchina Agricultural University, China; Tel:13662369316	yhwen@scau.edu.cn
64	Yannong Xiao	Prof.	College of Plant Science and Technology of Agricultural University, China; Tel:13098836377	xiaoyannong@yahoo.com.cn
65	Yinong Li	Prof.	Shenzhen Entry-Exit Inspection and Quarantine Bureau, China; Tel:13530289148	li_yinong@126.com
66	Yong Liu	Prof.	Institute of Plant Protection, Hunan Province, China; Tel:13307312011	haoasliu@163.com
67	Yuxi Duan	Prof.	Shenyang Agricultural University,China; Tel:13998253910	duanyx6407@163.com
68	Zhenchuan Mao	Asso Prof.	Institute of Vegetables and Flowers, Chinese Academy of Agricultural Sciences, China; Tel:15910897588	maozhenchuan@yahoo.com.cn
69	Zhong Ding	Asso Prof.	College of Plant Protection, Hunan Agricultural University, China; Tel:13875813230	dingzx88@yahoo.com.cn
70	Langyun Zhou	Miss	Syngenta (China) Investment Company	Annie.Zou@SYNGENTA.COM
71	Julian Chen	Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China	jlchen@ippcaas.cn
72	Dewen Qiu	Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China	dwqiu@ippcaas.cn
73	Jie Zhang	Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China	jzhang@ippcaas.cn
74	Xueping Zhou	Prof.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China	zzhou@zju.edu.cn

Volunteers

	Name	Title	Mailing Address	Email-address
75	ChoCho Htay	Ph.D Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:13269762868	2913600048@qq.com
76	Duqing Wu	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:18310985297	wuduqingsweet@sina.com
77	Fen Qiao	Ph.D Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:62815611	qiaofen121@163.com
78	Gaofeng Wang	Ph.D student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:13466708967	jksgo@126.com
79	Huan Peng	Dr.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:62815611	foumer@126.com
80	Jiangkuan Cui	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:18810600644	jk_cui@163.com
81	Jing Liu	Ph.D Stud	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China;	liujing3878@sina.com
82	Lilian Luo	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:18202740100	imluoliha@163.com
83	Linfeng Zhu	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:15001338196	zhulinfeng1234@126.com
84	Maoyan Liu	student	Hunan Argicultural University China; Tel:13272014266	82559204@qq.com
85	Qingsong Wu	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:13261038237	aaawqswqs@163.com
86	Shiqi Ou	Dr.	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:13520642383	jlccosq@126.com
87	Shujie Luo	Ph.D Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China; Tel:18501190531	702176115@qq.com
88	Ting Li	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China	915188700@qq.com
89	Xin Li	Student	Institute of Plant Protection, Chinese Academy of Agricultural Sciences, China, Huazhong Agricultural University	245914504@qq.com
90	Yu Meng	student	Jilin Argicultural University China; Tel:13661084716	1649825764@qq.com
91	Yuepeng Sun	student	Jilin Argicultural University; China;Tel:15613658082	316692672@qq.com

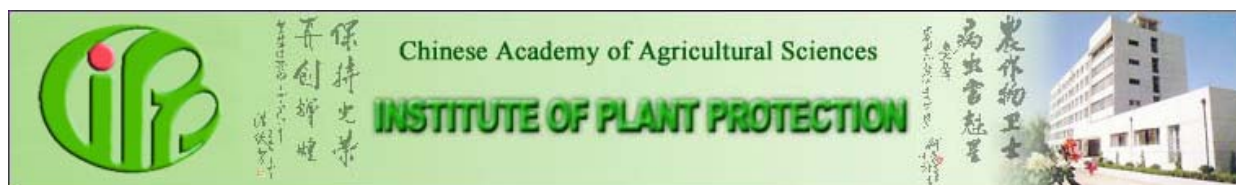
Map of Friendship Hotel and Beijing



北京友谊宾馆示意图

Plan of Beijing Friendship Hotel





Introduction of IPP, CAAS

The Institute of Plant Protection (IPP), Chinese Academy of Agricultural Sciences (CAAS), which was founded in 1957, is a non-profit national institute dedicated to both basic and applied sciences of plant protection in China. The mission of the institute is to study and seek the resolution to the main problems theoretically and economically important in plant protection of the country. IPP is committed to develop and extend the research achievements in plant protection, promote the agro-ecosystem and environmental protection, and develop the international collaboration and exchange.

In the institute there are three research departments of plant diseases, insect pests and pesticide, three administration sectors for scientific research management and international cooperation, general affairs, and human resources, a supporting sector of service center, an affiliated experimental station located in Langfang, Hebei Province. The national key laboratory for biology of plant diseases and pests, the key laboratory of pesticide chemistry and application technology of Ministry of Agriculture (MOA), biosafety research center (MOA), supervision and test for plant disease and pest resistance (MOA), and Chinese Society of Plant Protection are all based in the institute. IPP currently has 210 employees. IPP is one of the institutes certified by the Academic Degree Committee of the National Council to authority to grant doctoral degrees in plant pathology, agricultural entomology, pesticide science, weed science, agricultural microbiology, monitoring and forecasting of plant pests and biosafety.

IPP has been undertaking lots of research projects from the national program of plant protection, serving as a nationwide coordinator for the research collaboration in plant protection nationwide. Approximately 50 national and international research projects are under implementation, including the National Basic Research and Development Program, the National High-Tech Research and Development Program, the National Key Technological Research and Development Program, the projects of Natural Science Foundation of China, the ministerial and provincial research projects and international collaboration program.

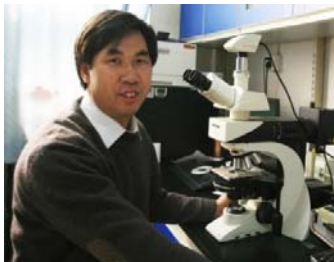
In IPP, more than 2500 scientific research papers and over 100 books have been published, including “Crop Diseases and Pests in China” and “Study on *Helicoverpa armigera*”. The institute has won more than 210 awards for scientific achievement, including 2 natural sciences awards, 2 national invention awards, 23 national advanced science awards, and 125 awards of the ministerial or provincial levels. Most of the researches have already been well recognized worldwide. The monographs “Epidemic systems of wheat stripe rust in China” and “Migration regularity and forecasting of armyworm moth” won second and third prizes in the National Natural Science Award respectively, and “Development of transgenic wheat resistant to barley yellow dwarf virus using genetic engineering approach” is one of the top ten achievements for science and technology in China in 1995. IPP hosts the 15th International Plant Protection Congress in 2004.

The institute won the first place for the contribution to scientific achievements in the comprehensive evaluation of nationwide agricultural institutions organized by MOA in 1992 and 1996. The national key laboratory for biology of plant diseases and pests was placed the top ten national laboratories in the 1st round evaluation of NSFC entrusted by National Planning Committee in 1996. The laboratory ranked 14th place among 56 key laboratories in life science in the 2nd round evaluation organized by NSFC entrusted by Ministry of Science and Technology in 2001. The ministerial key laboratory of pesticide chemistry and application technology was placed the top ten laboratories in the evaluation organized by MOA in 2002.

With the increasing progress of scientific research, improvement of working conditions and development of international collaboration, IPP will certainly be one of the national institutes with the majors covering main research fields in plant protection, the excellent capability to undertake the research projects and make the outstanding research progress. IPP will make fundamental, strategic and profound contributions to plant protection, agricultural production and food safety of the country.

For more information, please visit the website: <http://www.ipzcaas.cn/sites/IPP/ippc/ipzcaas>

Introduction of Nematology Laboratory, IPPCAAS



Principal Investigator: **De-Liang Peng** Dr. & Prof.
State Key Laboratory for Biology of Plant Diseases and Insect Pests,
Institute of Plant Protection, Chinese Academy of Agricultural Sciences
Yuanmingyuan West Road No.2, Beijing 100193, China
Tel: 86-10-62815611; Fax: 86-10-62896114
Email: dlpeng@ippcaas.cn pengdeliang@caas.cn

Research Areas

We pay more attention to the important agricultural nematodes, such as cyst nematodes (*H. avenae*, *H. filipjevi*, *H. glycines*, *H. elachista*), root-knot nematodes, root lesion nematodes, stem nematodes and quarantine nematodes. The research areas mainly focus on:

1. Nematode detection, early warning and risk analysis
2. Identification and functional analysis of parasitism/pathogenic effectors genes
3. Germplasm resistant identification and evaluation
4. Molecular interaction between nematode and host
5. Biological control and integrated nematode management
6. Environments biosafety assessment of transgenic soybean



Selected Publications

1. Peng H, Gao BL, Kong LA, Yu Q, Huang WK, He XF, Long HB, Peng DL*. Exploring the host parasitism of the migratory plant-parasitic nematode *Ditylenchus destructor* by expressed sequence tags analysis. PLoS ONE 2013 8(7): e69579 (*corresponding author)
2. Long H, Peng DL*, Huang WK, Peng H, Wang GF. Molecular characterization and functional analysis of two new β -1,4-endoglucanase genes (*Ha-eng-2*, *Ha-eng-3*) from the cereal cyst nematode *Heterodera avenae*. Plant Pathology 2012;62:953- 960
3. Long HB, Peng H, Huang WK, Wang GF, Gao BL, Mones M, Peng DL*. Identification and molecular characterization of a new β -1,4-endoglucanase gene (*Ha-eng-1a*) in the cereal cyst nematode *Heterodera avenae*. European Journal of Plant Pathology 2012;134:391–400

Fundings

1. 973 program (Grant No. 2013 CB127502) from the Ministry of Sciences and Technology, PI: De-Liang Peng
2. Special Fund for Agro-scientific Research in the Public Interest (Grant No. 200903040), PI: De-Liang Peng
3. National Natural Science Foundation (6 individual funds)
4. Environments biosafety assessment for Transgenic crops

基因组学 Workflow





卡尤迪生物科技（北京）有限公司

公司简介：

卡尤迪生物科技（北京）有限公司是一家致力于研发、生产世界一流品牌的小型便携化的分子生物实验室仪器的高科技生物企业。卡尤迪也是中国第一家、世界第二家致力于研发手持式通用荧光定量PCR仪的生物公司。本公司具有仪器、试剂交叉开发能力，可以进行核酸检测颠覆性的系统设计。

公司目前主要有PCR仪系列、金属浴系列、组织研磨器系列和便携核酸分析系列这四大产品线。

超薄型PCR仪是现在市场上最小巧，最轻薄的PCR仪。机身厚度仅有4厘米，极大的节省了实验空间；人机友好的触摸屏设计，真正实现了傻瓜式操作；12V直流电源供电，可车载供电，是您室外实验的最佳助手。

制冷/加热恒温金属浴系列是一种先进的温控仪器，是市场中体积最小巧，价格计，方便实用的多样化模块，安全环保的低能耗电源等产品优点让卡尤迪金属浴成为了北京各大科研院校老师学生喜爱的实验室常规仪器之一。

G10/G20是市场上动力最强劲的手持式电动组织研磨器，充电后可连续使用最长达10小时。加强版G50提供更强劲的动力，LCD屏幕观测实时转速，并可通过按钮轻松调节所需转速，是各种DNA\RNA提取试剂盒的最佳伴侣，在各种生物样品组织（包括动物、植物）的研磨提取中为您提供愉悦轻松且高效的研磨体验。

BATT-3蓄电池盒可为所有的卡尤迪产品提供外接移动电源，使得野外和移动使用成为可能。



北京市海淀区创业中路36号5层

010-62977520

www.coyotebio.com